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dUTPase inhibitors

Field of the Invention

The present invention relates to pharmaceuticals active against parasite dUTPase and methods for treating parasitical infections, especially malaria, by administering such compounds.

Technical Background

Deoxyuridine triphosphate nucleotidohydrolase (dUTPase) E.C. 3.6.1.23 is an ubiquitous enzyme which hydrolyzes deoxyuridine triphosphate (dUTP) to deoxyuridine monophosphate (dUMP) and pyrophosphate, typically in the presence of magnesium ions. This reaction is thought to occur primarily to limit pools of intracellular dUTP in order to prevent significant uridine incorporation into DNA during replication and repair. A second role of dUTPase is to provide substrate (dUMP) for the de novo synthesis of thymidylate.

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Two groups of researchers, McIntosh et al., PNAS, 89:8020-8024 (1992) and Strahler et al., PNAS, 90:4991-4995 (1993), have reportedly isolated the trimeric human dUTPase enzyme and characterized the enzyme by its cDNA and amino acid sequences.

20 McIntosh reported a cDNA of 526 base pairs containing an ORF which encoded a protein of 141 amino acids and a 3f flanking sequence following the ORF. Strahler reported the identical cDNA and amino acid sequence as did McIntosh, with the exception of two additional bases at the 51 end of the cDNA and a longer 3f flanking sequence. The human dUTPase reported by both groups was found to have a high degree of homology with dUTPase from other organisms including that from yeasts, bacteria and viruses. Strahler further reported that human dUTPase exists in both, phosphorylated and a non-phosphorylated forms.

International patent application no WO97/36916 discloses the sequence of nuclear and mitochondrial isoforms of dUTPase.

In both prokaryotic and eukaryotic cell systems, dUTPase has been clearly shown to be an essential enzyme, without which the cell will die. Lack of dUTPase leads to elevated cellular dUTP pools, resulting in an increased misincorporation of uridine into DNA. In **WO** 2005/066160

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addition to prokaryotes and eukaryotes, a number of viruses, such as herpes simplex, are known to encode a dUTPase function.

International patent application no WO95/15332 proposes a range of uridine di- and triphosphate analogues in which the oxygen atoms between phosphate groups are replaced with methylene, secondary amine or tertiary amine, and/or oxo functions on the phosphate are replaced with sulphur. These compounds are postulated as cytostatics for use against rapidly growing cancer cells and/or antivirals against herpes. Substantially similar compounds are disclosed in Zalud et al Adv. Exp.Med Biol. 1995 370 135-138 and Persson et al Biolog Med Biochem 1996 4 553-556. It should be noted, however that these compounds have been primarily designed for crystallographic purposes and the analysis of enzyme kinetics. These compounds therefore do not possess physicochemical attributes suggestive of a drug.

The present inventors have established that the substrate specificity of the dUTPases of certain protozoal and bacterial parasites of man differ from the corresponding human cellular and mitochondrial enzymes to such an extent that a specific set of inhibitor compounds can be prepared which selectively inhibit the parasite dUTPase without substantially inhibiting the human counterparts. Examples of such parasites include Plasmodium species especially P.falciparum responsible for malaria, Mycobacterial species, especially M tuberculosum responsible for tuberculosis and Leishmania spp.

Hidalgo-Zarco and González-Pacanowska Current Protein and Peptide Science, 2001, 2, 389-397 describe the isolation and characterisation of trypanosomal dUTPases. In contrast to the trimeric form of dUTPase shared by human and malarial enzymes, the trypanosomal enzyme is a dimmer. Competitive Inhibition of Leishmania dUTPase was shown by the triphosphate substrate analogue α - β -imido-dUTP, whereas no inhibition of that parasite was apparent in the case of 5'-O-(4-4'-dimethoxytrity!)-2'-deoxyuridine.

30 Brief description of the Invention

In accordance with a first aspect of the invention there are provided use of deoxyuridine derivatives of the formula I, in the manufacture of a medicament for the treatment or

prophylaxis of parasitic infections, particularly plasmodium infections in mammals, including man.

$$\begin{array}{c} R6 \\ R7 - E - C_0 - C_3 - alkyl - D - C_0 - C_3 - alkyl \\ R8 \end{array}$$

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A is O, S or CH₂;

B is O, S or CHR³;

 R^1 is H, C_1 - C_5 alkyl, C_2 - C_5 alkenyl, C_2 - C_5 alkynyl or a 5 or 6 membered, saturated or unsaturated ring containing 0 to 3 heteroatoms selected from N, O and S, the alkyl,

10 alkenyl, alkynyl or ring is optionally substituted with R⁴;

R² is H or F;

R³ is H, F, OH, NH₂ or a pharmaceutically acceptable ester, amide or ether thereof; or R² and R³ together form a chemical bond;

D is -NHCO-, -CONH-, -O-, -C(=O)-, -CH=CH, -C=C-, -NR⁵-

R⁴ is independently hydrogen, halo, cyano, amino, nitro, carboxy, carbamoyl, hydroxy, oxo, C₁-C₅ alkyl, C₁-C₅ haloalkyl, C₁-C₅ alkyloxy, C₁-C₅ alkanoyl, C₁-C₅ alkanoyloxy, C₁-C₅ alkylthio, -N(C₀-C₃-alkyl)₂, hydroxymethyl, aminomethyl, carboxymethyl; -SO₂N(C₀-C₃-alkyl), -SO₂C₁-C₅-alkyl, where n is 1 or 2;

R⁵ is H, C₁-C₃-alkyl, C₁-C₃-alkanoyl;

20 E is Si or C;

R⁶, R⁷ and R⁸ are independently selected from C₁-C₈ alkyl, C₂-C₈ alkenyl, C₂-C₈ alkynyl, or a stable monocyclic, bicyclic or tricyclic ring system which is saturated or unsaturated in which each ring has 0 to 3 heteroatoms selected from N, O and S, wherein any the R⁶, R⁷ and/or R⁸ group may be optionally substituted with R⁴;

25 and pharmaceutically acceptable salts thereof.

According to a second aspect of the invention there are provided novel compounds of the formula II

$$\begin{array}{c} R6 \\ R7-E-C_0-C_3-alkyl-D-C_0-C_3-alkyl-A \\ R8 \end{array}$$

where

5 A is O, S or CH₂;

B is O, S or CHR3;

R¹ is H, C₁-C₅ alkyl, C₂-C₅ alkenyl, C₂-C₅ alkynyl or a 5 or 6 membered, saturated or unsaturated ring containing 0 to 3 heteroatoms selected from N, O and S, the alkyl, alkynyl or ring is optionally substituted with R⁴;

10 R^2 is H or F;

R³ is H, F, OH, NH₂ or a pharmaceutically acceptable ester, amide or ether thereof; or R² and R³ together form a chemical bond;

D is -NHCO-, -CONH-, -O-, -C(=O)-, -CH=CH, -C=C-, -NR5-,

R4 is independently hydrogen, halo, cyano, amino, nitro, carboxy, carbamoyl, hydroxy,

oxo, C₁-C₅ alkyl, C₁-C₅ haloalkyl, C₁-C₅ alkyloxy, C₁-C₅ alkanoyl, C₁-C₅ alkanoyloxy, C₁-C₅ alkylthio, -N(C₀-C₃-alkyl)₂, hydroxymethyl, aminomethyl, carboxymethyl; -SO_nN(C₀-C₃-alkyl), -SO_nC₁-C₅ alkyl, where n is 1 or 2;

R⁵ is H, C₁-C₃ alkyl, C₁-C₃ alkanoyl;

E is Si or C;

20 R⁶ and R⁷ are independently selected from a stable monocyclic, bicyclic or tricyclic ring system which has an aromatic nature in which each ring has 0 to 3 heteroatoms selected from N, O and S;

R⁸ selected from C₁-C₈ alkyl, C₂-C₈ alkenyl, C₂-C₈ alkynyl, or a stable monocyclic, bicyclic or tricyclic ring system which is saturated or unsaturated in which each ring has

0 to 3 heteroatoms selected from N, O and S; wherein R⁶, R⁷ and R⁸ group are optionally substituted with R⁴;

with the proviso that if the group C_0 - C_3 alkyl-D- C_0 - C_3 alkyl is -O- CH_2 -, then the group $E(R^6)(R^7)(R^8)$ is not trityl, methoxylated trityl or tert butyldiphenylsilyl; or a pharmaceutically acceptable salt thereof.

- The novel compounds of the invention are useful in methods for the treatment or propylaxis, or in the manufacture of a medicament for such treatment or prophylaxis, of parasitic infections, such as Leishmania, trypansoma, human African trypanosomiasis, Chagas disease or plasmodium (malaria).
- The potency and selectivity of the compounds and methods of the invention, which presuppose substantial lipophilicity at the 5' position is surprising bearing in mind that the active site of the dUTPase enzyme is intended to recognize and accommodate highly polar, hydrophilic moieties, ie the triphosphorylated nucleotides.
- 15 Conveniently, A is -O- and B is -CHR³- thus defining a 2'-deoxyribose analogues.
 - Alternative preferred variants include those where A is -O- and B is -O-, or -S-, thus defining a dioxolane or especially an oxathiolane derivative.
- Other preferred variants include those wherein R² and R³ form a chemical bond and A is -O-, thus defining a 2'3'-dideoxy, didehydroribose derivative or R² and R³ are H, thus defining a 2',3'-dideoxyribose derivative.
- Still further preferred variants include those wherein R² and R³ form a chemical bond and A is -CH₂-, thus defining a 2-cyclopentene derivative or those wherein R² and R³ are H, thus defining a cyclopentane derivative.
- It is currently preferred that R³ is H, NH₂, OH or F. An alternative, but currently less favoured, R³ is a lipophilic ester such as straight or branched chain alkyl or benzyl ester or an ether such as straight or branched chain alkyl or benzyl ether or alkylated silyl function.
 - R¹ is preferably a small substituent, most preferably H.

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Favoured C₀-C₃-alkylene-D-C₀-C₃-alkylene configurations include aminomethylene, aminoethylene and aminopropylene, methylaminomethylene, methylaminoethylene, ethylaminomethylene, -(N-methyl)aminomethylene, -(N-methyl)aminopropylene and methyl-(N-methyl)aminomethylene. Currently the most preferred is -aminomethylene-. The order of the hetero atom and alkylene moieties in the indicated groups as used herein corresponds to the configuration of Figure I or II as depicted above, that is "aminomethylene" has the nitrogen atom adjacent E and the methylene moiety proximal to the base.

Particularly preferred C₀₋C₃-alkylene-D-C₀-C₃-alkylene configurations include -O-,
oxymethylene, oxyethylene, oxypropylene methyloxymethylene and methyloxyethylene.
Currently the most preferred in this series is -oxymethylene-.

Preferably at least one of R⁶, R⁷ and/or R⁸ has an aromatic nature, although this tends to be less important if R³ has a lipophilic nature. Conveniently two of R⁶, R⁷ and/or R⁸ have an aromatic nature and the invention even embraces compounds wherein all three have an aromatic nature.

Ring systems for R⁶, R⁷ and/or R⁸ are typically bonded direct to E, but may optionally be bonded to E via a methylene linker. For example R⁶ may be optionally substituted benzyl, thereby representing phenyl bonded through a methylene to E.

Ring systems for R⁶, R⁷ and/or R⁸ having an aromatic nature include optionally substituted heteroaryls such as furyl, thienyl, pyrrolyl, pyrrazolyl, imidazolyl, pyridyl, pyrimidinyl, pyridazinyl, oxazolyl, isoxazolyl, thizolyl, isothiazolyl, especially pyridyl; and optionally substituted carbocycles such as phenyl. Ring systems having an aromatic nature also include multi-ring systems wherein only one ring has an aromatic nature such as indolinyl and ring systems wherein more than one ring has an aromatic nature such as naphthyl or any of the above heterocyclic rings fused to phenyl, such as benzimidazolyl.

Convenient values for R⁶, R⁷ and/or R⁸ include heterocyles such as furyl, thienyl, pyranyl, pyrrolyl, pyrrolidinyl, pyrazolyl, pyrazolyl, pyrazolidinyl, imidazolyl, imidazolidinyl, pyridyl, piperidinyl, pyrazinyl, piperazinyl, pyrimidinyl, pyridazinyl, oxazolyl, oxazolidinyl, isoxazolyl, isoxazolyl, isoxazolidinyl, morpholinyl, thiazolyl, thiazolyl, isothiazolyl, isothiazolyl, especially pyridyl, and carbocycles such as

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cycloalkyl, cycloalkenyl and especially phenyl. Alternative values for R⁶, R⁷ and/or R⁸ include straight or branched alkyl, including methyl, ethyl, i-propyl and t-butyl.

The optional substituent(s) to R⁶, R⁷, and/or R⁸ include 1 to 3, preferably 1 substituent per ring, selected from halo, preferably fluoro, cyano (preferably cyano), amino, nitro, carboxy, carbamoyl, hydroxy, oxo, C₁-C₅ alkyl, preferably methyl or t-butyl, C₁-C₅ haloalkyl, preferably triflouromethyl, C₁-C₅ alkyloxy, preferably methoxy, C₁-C₅ alkanoyl, preferably acetyl, C₁-C₅ alkanoyloxy, preferably acetoxy, C₁-C₅ alkylthio, -N(C₀-C₃-alkyl)₂, preferably NHMe or NMe, hydroxymethyl, aminomethyl, carboxymethyl; - SO_nN(C₀-C₃-alkyl) (n=1, 2), preferably SO₂NH₂ or SO₂NMe₂ or -SO_nC₁-C₅-alkyl, (n=1,2) preferably sulphonylmethyl or sulphinylmethyl.

Favoured $R^6(R^7)(R^8)$ -E- configurations include -C(Ph)₃ (trityl), -CH(Ph)₂, -CH₂Ph, -Si(Ph)₂(t-Bu), 1,1-bis(4-methylphenyl)-1'-pyrenylmethyl, where Ph is phenyl or phenyl substituted with R^4 .

Note, however, that the novel compounds of the invention exclude by way of proviso certain compounds with common protecting groups at the 5'-oxygen of the nucleoside, such as 5'-O-trityl, methoxylated 5'-O-trityl or 5'-O-tert.butyldiphenylsilyl. Accordingly 5'-O-(4',4'-dimethoxytrityl)-2'-deoxyuridine is outside the scope of the novel compound aspect of the invention. This exclusion of trityl and tBuPh₂Si in the compound claims only is not believed to be required in respect of other permutations of C_O-C₃alkyl-D-C₀-C₃ alkyl, such as compounds wherein D is N. The novel compounds of the invention will however typically avoid conventional hydroxyl protecting groups (such as those cited in Greene below), when C_O-C₃alkyl-D-C₀-C₃ alkyl is -O-CH₂-. It will be appreciated, however, that the use/method of treatment aspects of the invention include those compounds excluded from the compound claims by proviso.

Compounds wherein E is carbon are currently favoured on pharmacokinetic grounds, although compounds with E as Si have shown advantageous potency and selectivity.

The compounds of the invention include a number of chiral centres, and the invention extends to include racemates, enantiomers and stereoisomers at each of these centres. For example the ring carbon attached to the uracil N1 in Formula I may be in the alpha

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(down) or preferably the beta (up) configuration. R_2 as F in Formula I may be in the ribo (down) position although it is currently preferred to have the arabino (up) position. It is currently preferred that the ring carbon intermediate A and B in Formula I projects the adjacent C_0 - C_3 alkylene in the beta configuration.

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Compounds of the invention are generally at least 80% preferably at least 90% such as 97% stereoisometrically pure at chiral centres.

Additional aspects of the invention include a pharmaceutical composition comprising a compound of the formula I in conjunction with a pharmaceutically acceptable carrier or diluent therefore. The invention further provides a method for the treatment or prophylaxis of parasite infections, such as malaria, tuberculosis or leishmaniasis, in man or a zoonose vector comprising the administration of an effective amount of a compound of the formula I to a patient in need thereof, or to the vector.

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While it is possible for the active agent to be administered alone, it is preferable to present it as part of a pharmaceutical formulation. Such a formulation will comprise the above defined active agent together with one or more acceptable carriers or excipients and optionally other therapeutic ingredients. The carrier(s) must be acceptable in the sense of being compatible with the other ingredients of the formulation and not deleterious to the recipient.

The formulations include those suitable for rectal, nasal, topical (including buccal and sublingual), vaginal or parenteral (including subcutaneous, intramuscular, intravenous and intradermal) administration, but preferably the formulation is an orally administered formulation. The formulations may conveniently be presented in unit dosage form, e.g. tablets and sustained release capsules, and may be prepared by any methods well known in the art of pharmacy.

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Such methods include the step of bringing into association the above defined active agent with the carrier. In general, the formulations are prepared by uniformly and intimately bringing into association the active agent with liquid carriers or finely divided solid carriers or both, and then if necessary shaping the product. The invention extends to methods for preparing a pharmaceutical composition comprising bringing a

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compound of Formula I or its pharmaceutically acceptable salt in conjunction or association with a pharmaceutically acceptable carrier or vehicle. If the manufacture of pharmaceutical formulations involves intimate mixing of pharmaceutical excipients and the active ingredient in salt form, then it is often preferred to use excipients which are non-basic in nature, i.e. either acidic or neutral.

Formulations for oral administration in the present invention may be presented as discrete units such as capsules, cachets or tablets each containing a predetermined amount of the active agent; as a powder or granules; as a solution or a suspension of the active agent in an aqueous liquid or a non-aqueous liquid; or as an oil-in-water liquid emulsion or a water in oil liquid emulsion and as a bolus etc.

With regard to compositions for oral administration (e.g. tablets and capsules), the term suitable carrier includes vehicles such as common excipients e.g. binding agents, for example syrup, acacia, gelatin, sorbitol, tragacanth, polyvinylpyrrolidone (Povidone), methylcellulose, ethylcellulose, sodium carboxymethylcellulose, hydroxypropylmethyl cellulose, sucrose and starch; fillers and carriers, for example corn starch, gelatin, lactose, sucrose, microcrystalline cellulose, kaolin, mannitol, dicalcium phosphate, sodium chloride and alginic acid; and lubricants such as magnesium stearate, sodium stearate and other metallic stearates, stearic acid, glycerol stearate, silicone fluid, talc waxes, oils and colloidal silica. Flavouring agents such as peppermint, oil of wintergreen, cherry flavouring or the like can also be used. It may be desirable to add a colouring agent to make the dosage form readily identifiable. Tablets may also be coated by methods well known in the art.

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A tablet may be made by compression or moulding, optionally with one or more accessory ingredients. Compressed tablets may be prepared by compressing in a suitable machine the active agent in a free flowing form such as a powder or granules, optionally mixed with a binder, lubricant, inert diluent, preservative, surface-active or dispersing agent. Moulded tablets may be made by moulding in a suitable machine a mixture of the powdered compound moistened with an inert liquid diluent. The tablets may be optionally be coated or scored and may be formulated so as to provide slow or controlled release of the active agent.

Other formulations suitable for oral administration include lozenges comprising the active agent in a flavoured base, usually sucrose and acacia or tragacanth; pastilles comprising the active agent in an inert base such as gelatin and glycerin, or sucrose and acacia; and mouthwashes comprising the active agent in a suitable liquid carrier. Dosages are set in the conventional manner to take into account the severity of the disease, the susceptibility of the parasite strain, the size and metabolic health of the patient, the mode and form of administration, concomitant medication and other relevant factors. The compounds of the invention may be administered at a daily dose generally in the range 0.1 to 200 mg/kg/day, advantageously, 0.5 to 100 mg/kg/day, more preferably 10 to 50mg/kg/day, such as 10 to 25 mg/kg/day. A typical dosage rate for a normal adult will be around 50 to 500 mg, for example 300 mg, once or twice per day.

The compounds of formula I can form salts which form an additional aspect of the invention. Appropriate pharmaceutically acceptable salts of the compounds of formula I include salts of organic acids, especially carboxylic acids, including but not limited to acetate, trifluoroacetate, lactate, gluconate, citrate, tartrate, maleate, malate, pantothenate, isethionate, adipate, alginate, aspartate, benzoate, butyrate, digluconate, cyclopentanate, glucoheptanate, glycerophosphate, oxalate, heptanoate, hexanoate, fumarate, nicotinate, palmoate, pectinate, 3-phenylpropionate, picrate, pivalate, proprionate, tartrate, lactobionate, pivolate, camphorate, undecanoate and succinate, organic sulphonic acids such as methanesulphonate, ethanesulphonate, 2-hydroxyethane sulphonate, camphorsulphonate, 2-napthalenesulphonate, benzenesulphonate, p-chlorobenzenesulphonate and p-toluenesulphonate; and inorganic acids such as hydrochloride, hydrobromide, hydroiodide, sulphate, bisulphate, hemisulphate, thiocyanate, persulphate, phosphoric and sulphonic acids.

Examples of monocyclic rings for R¹ include heterocycles such as furyl, thienyl, pyranyl, pyrrolyl, pyrrolinyl, pyrrolinyl, pyrazolyl, pyrazolyl, pyrazolinyl, pyrazolidinyl, imidazolyl, imidazolidinyl, pyridyl, piperidinyl, pyrazinyl, piperazinyl, pyrimidinyl, pyridazinyl, oxazolyl, oxazolidinyl, isoxazolyl, isoxazolyl, isoxazolyl, morpholinyl, thiazolyl, thiazolyl, isothiazolyl, isothiazolidinyl, especially pyridyl, and carbocycles such as cycloalkyl, cycloalkenyl and phenyl.

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Examples of monocyclic, bicyclic or tricyclic rings for R⁶, R⁷ and/or R⁸ include heterocycles such as furyl, thienyl, pyranyl, pyrrolyl, pyrrolinyl, pyrrolinyl, pyrrolinyl, pyrrazolyl, pyrazolyl, pyrazolinyl, pyrazolidinyl, imidazolyl, imidazolinyl, imidazolidinyl, pyridyl, piperidinyl, pyrazinyl, piperazinyl, pyrimidinyl, pyridazinyl, oxazolyl, oxazolidinyl, isoxazolyl, thiazolyl, thiazolyl, thiazolyl, isothiazolyl, isothiazolyl, isothiazolyl, thiadiazolyl, triazolyl, and the like or bicyclic rings especially of the above fused to a phenyl ring such as indolyl, quinolyl quinolyl, isoquinolinyl, benzimidazolyl, benzothiazolyl, benzothiazolyl, benzothiazolyl, benzothiazolyl, benzothiazolyl, benzothiazolyl, benzothiazolyl, benzothianyl, 9-(9-alkyl)xanthenyl, 9-(9-alkyl)xanthenyl, 9-(9-alkyl)xanthenyl, 9-heteroarylxanthenyl, 9-(9-heteroaryl)xanthenyl, 9-(9-heteroaryl)xanthenyl, 9-(9-heteroaryl)xanthenyl, 5-(5-heteroaryl)fluorenyl) and the like.

Examples of monocyclic, bicyclic or tricyclic ring systems with an aromatic nature for R⁶,and/or R⁷ include heteroaryls such as furyl, thienyl, pyrrolyl, pyrazolyl, imidazolyl, pyridyl, pyrazinyl, pyrimidinyl, pyridazinyl, oxazolyl, isoxazolyl, thiazolyl, isothiazolyl, thiadiazolyl, tetrazolyl, triazolyl, and the like or bicyclic rings especially of the above fused to a phenyl ring such as indolyl, quinolyl, quinolinyl, isoquinolinyl, benzimidazolyl, benzothiazolyl, benzoxazolyl, benzotriazolyl, benzofuryl, benzothienyl etc. Additional rings include xanthenyl (such as 9-xanthenyl, 9-alkylxanthenyl, 9-(9-alkyl)xanthenyl, 9-phenylxanthenyl, 9-(9-phenyl)xanthenyl, 9-heteroarylxanthenyl, 9-(9-heteroaryl)xanthenyl), dibenzosuberyl, 5-dibenzosuberyl, fluorenyl (such as 5-fluorenyl, 5-(5-alkyl)fluorenyl, 5-(5-phenyl)fluorenyl, 5-(5-heteroaryl)fluorenyl) and the like.

Examples of carbocycles for R⁶, R⁷ and/or R⁸ include monocyclic rings such as phenyl, cyclohexenyl, cyclohexanyl, cyclopentanyl, bicyclic rings such as indanyl, napthyl, and tricyclic rings such as adamantyl, and the like.

The carbo or heterocyclic ring may be bonded via a carbon or via a hetero atom,
typically a nitrogen atom, such as N-piperidyl, N-morpholinyl etc. Other examples of
such ring systems may also be found in J. Fletcher, O. Dermer, R. Fox, Nomenclature
of Organic Compounds, pp. 20-63 (1974).

The term "C₁-C₅ alkyl" includes such groups as methyl, ethyl, n-propyl, isopropyl, n-butyl, s-butyl, t-butyl, cyclopropyl, n-pentyl and the like with C₁-C₈ alkyl further including n-hexyl, 3-methylpentyl, and the like.

- 5 The term "halo" and "halogen" refer to chloro, bromo, iodo, and especially fluoro.
 - "C₁-C₅ alkoxy" refers to those groups such as methoxy, ethoxy, propoxy, t-butoxy and the like.
- "C₂-C₅ alkenyl" refers to those groups such as vinyl, 1-propen-2-yl, 1-butene-4-yl, 1-pentene-5-yl, 1-butene-1-yl and the like, with C₂-C₈ alkenyl further including hex-3-enyl and the like.
- "C₁-C₅ alkylthio" refers to those groups such as methylthio, ethylthio, t-butylthio, and the like.
 - "C₁-C₅ alkanoyl" refers to groups such as acetyl, propionyl, butyryl and the like.
- "C₁-C₅ alkanoyloxy" refers to those groups such as acetoxy, propionoxy, formyloxy, butyryloxy, and the like.
 - The term "C₂-C₈ alkenoxy" includes groups such as ethenyloxy, propenyloxy, iso-butoxy ethenyl, and the like.
- The term "C₂-C₅ alkynyl" includes groups such as ethynyl, propynyl, butynyl, pentynyl, and the like with C₂-C₈ alkynl further including hexynyl and the ike.

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- The term "halo C₁-C₅ alkyl" includes alkyls substituted 1, 2 or 3 times by a halogen including groups such as trifluoromethyl, fluoromethyl, 2-dichloroethyl, 2,2-difluoroethyl, 2,2-trifluoroethyl, 3,3-difluoropropyl, 1,1-2,2,2 pentafluoroethyl and the like.
- The term $-C_0-C_3$ -alkylene- as a bivalent in expressions such as $-C_0-C_3$ -alkylene-D-C₀-C₃-alkylene includes a bond (i.e C₀), methylene (C₁), ethylene (C₂), 1,1-dimethyl-

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methylene (C_3), propylene (C_3) and the like, with each $-C_0$ - C_3 -alkylene- being selected independently.

The term (C_0 - C_3 -alkyl) in monovalent expressions includes H (i.e C_0), Me (C_1), Et (C_2), propyl (C_3) with each C_0 - C_3 -alkyl being selected independently. Accordingly -N (C_0 - C_3 -alkyl)₂ includes -NH₂, - NHMe, NHEt NHPr, -N(Me)₂, N(Et)₂ etc, -SO₂N(C_0 - C_3 -alkyl)₂, includes -SO₂NH2, -SO₂NHMe, -SO₂N(Me)₂ etc

As used herein, "the esters, amides and ethers thereof" refer to the appropriate derivatives of each of the preceding hydroxyl or amino groups in the respective definition.

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Representative esters thus include carboxylic acid esters in which the non-carbonyl moiety of the carboxylic acid portion of the ester grouping is selected from straight or branched chain alkyl (for example, methyl, n-propyl, t-butyl, or n-butyl), cycloalkyl, alkoxyalkyl (for example, methoxymethyl), araikyl (for example benzyl), aryloxyalkyl (for example, phenoxymethyl), aryl (for example, phenyl, optionally substituted by, for example, halogen, C₁₋₄ alkyl, or C₁₋₄ alkoxy) or amino); sulphonate esters, such as alkylor aralkylsulphonyl (for example, methanesulphonyl); amino acid esters (for example, L-valyl or L-isoleucyl); and mono-, di-, or tri-phosphate esters. In such esters, unless otherwise specified, any alkyl moiety present advantageously contains from 1 to 18 carbon atoms, particularly from 1 to 6 carbon atoms, more particularly from 1 to 4 carbon atoms. Any cycloalkyl moiety present in such esters advantageously contains from 3 to 6 carbon atoms. Any aryl moiety present in such esters advantageously comprises an optionally R⁴-substituted phenyl group.

Pharmaceutically acceptable esters thus include C₁-C₂₂ fatty acid esters, such as acetyl, t-butyl or long chain straight or branched unsaturated or omega-6 monounsaturated fatty acids such as palmoyl, stearoyl and the like.

Alternative aryl or heteroaryl esters include benzoyl, pyridylmethyloyl and the like any of which may be substituted with R⁴, Preferred pharmaceutically acceptable esters include aliphatic L-amino acid esters such as leucyl, isoleucyl and especially valyl. Additional preferred amino acid esters include the 2-O-AA-C₃-C₂₂ fatty acid esters described in

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WO99 09031, where AA is an aliphatic amino acid ester, especially those derived from L-lactic acid and L-valyl.

Pharmaceutically acceptable ethers include straight or branched chain saturated or omega 6 unsaturated C₁-C₂₂ alkyl ethers such as methyl ethers, t-butyl ethers or aryl or heteroaryl ethers such as phenoxy, benzylether, pyridylmethyl ether, any of which may be substituted with R⁴.

Alternative ethers include alkylated silyl functions such as -Si(C₁-C₅-alkyl)₃ such as -Si(t-Bu)(CH₃)₂, or -Si(Ph)₂(t-Bu), -C(Ph)₃ (trityl), -CH(Ph)₂, -CH₂Ph,1,1-bis(4-methylphenyl)-1'-pyrenylmethyl and the like.

Pharmaceutically acceptable amides include those derived from C_1 - C_2 branched or straight chain aminoalkyl optionally including 1 to 3 unsaturations and/or optionally substituted with R^4 , or anilines or benzylamines. Preferred amides include those formed from reaction of the amine with a C_1 - C_4 straight or branched chain alkanoic acid. Other pharmaceutically acceptable amides of amine functions of R^2 or R^{11} correspond to the esters indicated above.

20 It is currently preferred that the ester, amide or ether is lipophilic in nature.

Compounds of the invention are typically synthesized as outlined below.

Scheme 1 depicts a method for alkylation of the 5'-position of a nucleoside or a nucleoside analogue.

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Nucleoside derivative (1a) wherein A, B, R¹ and R² are as defined above for formula I and D is O or NH, can be reacted with an alkylating agent of formula 1b wherein R⁶, R⁷, R⁸ and E are as defined above for formula I and Lg is a leaving group that can be replaced by the nucleophile D, in a solvent like pyridine optionally in the presence of a catalyst such as dimethylaminopyridine or in a solvent like dimethylformamide in the presence of a catalyst like imidazole, to provide 5'-alkylated nucleoside analogues (1c). Various alkylating agents (1b) are available either commercially or in the literature, se for example Greene, "Protective Groups in Organic Synthesis (John Wiley & Sons, New York, 1981). For example, they can be prepared by transforming the hydroxy group of the corresponding alcohol into a leaving group such as a halide like chloride or bromide by treatment with a halogenating agent such as acetyl bromide or thionyl chloride or the like or they can be transformed into a derivative of sulfonic acid like a mesyl, tosyl, triflate or the like by treatment with for example the anhydride or acid chloride of the desired sulfonic acid derivative. An example of a route to alkylating agents is shown in scheme 2.

Scheme 2

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Reaction of an electrophilic carbonyl compound like a keto compound (2a) or any carboxylic acid derivative for instance an ester or acid halide, and a suitable nucleophile for example a Grignard reagent (2b) or an organolithium reagent, provides the alcohol (2c). The formed hydroxy group can subsequently be transformed into a leaving group as described above thus forming the alkylating agent (1b). Examples of the above procedure are described in the literature, se for example Hodges *et al.*, J. Org. Chem. 56, 1991, 449-452, and Jones *et al.*, J. Med. Chem. 33, 1990, 416-419.

Compounds wherein the leaving group in the alkylating agent (1b) is spaced by a C₁-C₃-alkylene chain, available either commercially or in the literature, may also be used as

alkylating agents in scheme 1. An example of a route to a compound containing a C_2 -alkylene chain is shown in scheme 3.

5 Scheme 3

A reaction performed with triphenylmethyl sodium (3a) and ethylene oxide provides alcohol (3b). Subsequent transformation of the hydroxy group into a leaving group for example as described above provides alkylating agent (3c). Use of any other appropriate electrophilic reagent for example formaldehyde, provides analogues with other length of the C₁-C₃-alkyl chain. Se for example Wooster *et al.*, J. Amer. Chem. Soc., 60, 1938, 1666 and McPhee *et al.*, J. Amer. Chem. Soc. 65, 1943, 2177, 2180. Alternatively, alkylating agents containing a C₁-C₃-alkyl chain may be obtained by reduction of an appropriate acyl derivative to the desired alcohol.

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A suitable acylating agent like the acid chloride or anhydride can be used to acylate the amino group of a 5'-aminonucleoside, thus providing compounds according to the general formula I where D is -CONH-.

The 5'-substituent can also be introduced by way of a Mitsunobu reaction of a desired alcohol and the 5'-unprotected nucleoside derivative as illustrated in scheme 3A.

R7-
$$E$$
-(CH₂)n-D-(CH

Scheme 3A

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Treatment of a desired optionally suitably protected alcohol (3Aa) and a nucleoside derivative (3Ab) with triphenyl phosphine and DIAD in a solvent like THF provides the nucleoside analogue (3Ac).

An example of the introduction of an ether group at the 3'-position of the nucleoside analogue is shown in scheme 4.

Scheme 4

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Treatment of a 5'-substituted nucleoside analogue (4a) with a silylating agent for example tert-butyldimethylsilyl chloride in a solvent like dimethylformamide in the presence of a catalyst like imidazole, provides 3'-O-silylated derivatives (4b).

- Other ether or ester groups can be introduced at the 3'-position by methods known in the art, for example by treating the 3'-OH nucleoside with the desired alkylating or acylating agent optionally in the presence of a suitable base, se for example Greene, "Protective Groups in Organic Synthesis (John Wiley & Sons, New York, 1981).
- Nucleoside analogues used in the synthesis of compounds according to the present invention are available either commercially or in the literature or they can be prepared as described herein. For example compound 1 wherein B is CH₂F, R¹ and R² are H and A and D are O *i.e.* FLU (3'-fluoro-2', 3'-dideoxyuridine) can be prepared in analogy with the procedure described for FLT (Balzarini et al. Blochem. Pharmacol. 37, 2847, 1988).

 The didehydro derivative d4U (2',3'-didehydro-2´,3'-dideoxyuridine) can be prepared in analogy with the procedure described for d4T (2',3'-didehydro-2´,3'-dideoxythymidine, Stavudine, Balzarini et al.; Mol. Pharmacol. 32, 162, 1987). 2'-Fluoro-2'deoxyarabino

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furanosyluracil is conveniently prepared for example as described by H. Howell in J. Org. Chem., 53, 85, 1988 and the corresponding ribo derivative, 2'-fluoro-2'deoxyribofuranosyluracil can be prepared as described for example by Mercer et al. in J. Med. Chem., 30, 670-675, 1987.

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5'-aminonucleoside analogues, useful for the preparation of compounds according to the general formula I wherein D is NH or -CONH- can be prepared from the corresponding 5'-alcohols by a displacement-reduction sequence for example as shown in scheme 5.

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$$HO \longrightarrow N \longrightarrow O \longrightarrow Ph_3P \longrightarrow N_3 \longrightarrow O \longrightarrow O \longrightarrow H_{20}Pd/C \longrightarrow O \longrightarrow Sc$$

Scheme 5

Nucleoside analogue 5a where Y is F or suitably protected OH or NH₂, can be reacted with triphenylphosphine in a solvent like carbon tetrabromide followed by displacement of triphenylphosphine oxide with azide ion to form 5b. Alternatively the hydroxy group can be transformed into a leaving group such as a halide like chloride or bromide or a derivative of sulphonic acid such as a tosyl, mesyl or a triflyl group which is subsequently displaced by azide ion. Reduction of the azide group for example by catalytic hydrogenation over palladium on carbon, gives the amino derivative 5c.

The procedures described in scheme 5 can be also applied to carbocyclic uridine and thiouridine analogues providing compounds useful for the preparation of compounds of the general formula I wherein A is CH₂ and S.

Detailed Description of the Embodiments

Various aspects of the invention, including end product inhibitors and intermediates towards those inhibitors will now be described by way of illustration only with reference to the following non-limiting examples. Note that the exemplified intermediates, such as the acyclic side chain building blocks are readily reacted with alternative bases to form additional compounds of the invention.

Example 1

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1-(5-Trityloxymethyl-2,5-dihydro-furan-2-yl)-1*H*-pyrimidine-2,4-dione or 5'-O-trityl-2',3'-dideoxydidehydrouridine (1)

2',3'-Dideoxy-didehydrouridine (0.30 g, 1.43 mmol) and triphenylmethyl chloride (0.44g, 1.57 mmol) was stirred in dry pyridine (10 ml) at 50 °C overnight under an atmosphere of nitrogen. The reaction mixture was poured into ice-H₂O (30 ml) with vigorous stirring and filtered. The precipitate was dissolved in EtOAc (50 ml) and the solution was washed with 0.5M HCl (20 ml), H₂O (20 ml), dried (Na₂SO₄) and reduced *in vacuo* to yield a crude product, which was purified by gradient flash column chromatography eluting with 0→3% MeOH/CHCl₃ to yield the title compound as white crystals (0.37 g, 58%).

¹H NMR (300MHz;MeOH): δ 3.56 (2H, m, 5'-H), 5.02 (1H, m, 4'-H), 5.08 (1H,m, 5-H), 5.93 (1H, m, 1'-H), 6.40 (1H, m, 2'-H), 7.09 (1H, m, 3'-H), 7.30-7.44 (15H, m, Ph-H), 7.87 (1H, d, J = 8.1 Hz, 6-H);

¹³C NMR (75MHz;MeOH): δ 64.82 (5'-CH₂), 86.38 (1'-CH), 87.84 (a), 90.04 (4'-CH), 102.70 (5-CH), 126.79 (Ph-CH), 127.79 (2'-CH), 128.41 (Ph-CH), 129.20 (Ph-CH), 134.89 (3'-CH), 141.79 (6-CH), 143.49 (Ph-C), 151.04 (2-C), 159.95 (4-C); MS (Cl/NH₃., m/z); 470.2 (M+NH₄⁺, 100%), 453.1 (M+H⁺, 20%),;

30 HRMS (ES+ve., M+H): Calculated for C₂₈H₂₄N₂O₄, requires 453.1814, found 453.1807.

IR_{vmax}/cm⁻¹ (KBr): 714 and 756 (Aromatic, monosubstituted), 1681.0 (C=O) and 1692.3 (C=O); Mp: 68°C, R_f (10% MeOH / CHCl₃): 0.30;

Example 2

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5'-O-tert-butyldiphenylsilyl-2',3'-dideoxydidehydrouridine (2)

2'3'-dideoxy-didehydrouridine (0.30 g, 1.43 mmol) in dry DMF (10ml) were added dropwise under an atmosphere of nitrogen, with ice bath cooling, to a stirred solution of *tert*-butyldiphenyllsilylchloride (0.41 ml, 1.57 mmol) and imidazole (0.21 g, 3.14 mmol) in dry DMF (10 ml). The mixture was allowed to warm to room temperature and stirred ovemight. H_2O (10 ml) was added and the mixture was extracted with $CHCl_3$ (2 x 30 ml). The combined extracts were washed with saturated aqueous $NaHCO_3$ solution (10 ml) and H_2O (10ml), dried (Na_2SO_4) and reduced *in vacuo* to obtain a crude product, which was purified by gradient flash column chromatography eluting with $0\rightarrow 3\%$ MeOH/CHCl₃ to yield the title compound as colourless viscous oil (0.46 g, 73%).

¹H NMR(300MHz;MeOH): δ 1.15 [9H, s, C(C*H*₃)₃], 3.95 (1H, dd, *J* = 2.9, 11.7 Hz, 5'-H), 4.06 (1H, dd, *J* = 2.9, 11.7 Hz, 5'-H), 4.97 (1H, m, 4'-H), 5.26 (1H, d, *J* = 8.1 Hz, 5-H), 5.58 (1H, m, 1'-H), 6.38 (1H, m, 2'-H), 7.10 (1H, m, 3'-H), 7.34-7.55 (6H, m, Ph-CH), 7.66-7.81 (5H, m, Ph-H and 6-H).

¹³C NMR(75MHz;MeOH): δ 19.81 [*C*(CH₃)₃], 26.99 and 27.42 [C(*C*H₃)₃], 65.40 (5'-CH₂), 87.56 (1'-CH), 90.06 (4'-CH), 102.96 (5-CH), 126.96 (2'-CH), 128.41 (Ph-CH), 128.32 (Ph-CH), 128.12 (Ph-CH), 130.03 (Ph-CH), 130.47 (Ph-CH), 130.59 (Ph-CH), 132.78 (Ph-C), 133.46 (Ph-C), 134.99 (3'-CH), 135.25 (Ph-CH), 135.79 (Ph-CH) 135.99 (Ph-CH), 141.20 (6-CH), 150.99 (2-C), 163.45 (4-C); MS (Cl/NH₃., *m/z*); 449.1 (M+H⁺, 50%), 466.2 (M+NH₄⁺, 100%); HRMS (ES+ve., M+H⁺): Calculated for C₂₅H₂₈N₂O₄Si, requires 449.1896, found 449.1894.

IR_{vmax}/cm⁻¹ (film): 1697.3 (C=O). R_f (10% MeOH / CHCl₃): 0.73.

Example 3

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Diphenyl (pyridin-3-yl) methanol (3)

A solution of 3-bromopyridine (10 g, 0.063 mol) in dry THF (200 mL)/hexane (50 mL) was cooled to -90 °C. To this cooled solution was added n-BuLi (2.2 M, 32 mL, 0.063 mol) slowly and allowed to stir for 30 min under N₂ atmosphere. A solution of benzophenone (11.5 g, 0.063 mol) in dry THF (50 mL) was added to this at the same temperature over a period of 30min. The reaction mixture was warmed slowly to RT and allowed to stir another 3h at RT. The reaction mixture was cooled, quenched with water (200 mL) and extracted with ethylacetate (2x100 mL). The organic layer was dried, concentrated under vacuum and crude purified by column chromatography over silica gel (30% ethyl acetate in petroleum ether) to give the title product (3.3 g).

Example 4

5'-O-Tosyl-2'-deoxyuridine (4)

To an ice-cold solution of 2'-deoxyuridine (5 g, 0.0219 mol) in dry pyridine (25 mL) tosyl chloride (5 g, 0.0263 mol) was added portion wise with stirring. The reaction mixture was stirred at 0 °C for 12h. The reaction mixture was concentrated under vacuum and the crude residue was washed with diethyl ether (5x25 mL). The residue was further treated with water. The solid precipitate formed was filtered, washed with water (2x25

mL), diethyl ether (5x25 mL) and petroleum ether (5x25 mL). The solid was dried under vacuum and used for next reaction without any purification. Yield: 7.5 g, 89%.

TLC: CHCl₃/MeOH, 4:1, R_f=0.6

5 Example 5

5'-Azido-2',5'-dideoxyuridine (5)

To a solution of 5'-O-tosyl-2'-deoxyuridine (13.5 g, 0.035 mol) in dry DMF (90 mL) was added NaN₃ (9.2 g, 0.141 mol) and the reaction mixture was allowed to stir at 95 °C for 12h. The reaction mixture was cooled, the solid residue was removed by filtration and the filtrate was concentrated under vacuum to give the crude product. The crude was purified by column chromatography on silica gel (4% methanol in chloroform) which gave the title product (5.2 g, 56%) as a white solid.

TLC: CHCl₃/MeOH, 4:1, R_f=0.45

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Example 6

5'-Amino-2',5'-dideoxyuridine (6)

To a mixture of 5'-azido-2',5'-dideoxyuridine (5 g, 0.0197 mol) in methanol/water (150 mL, 1:1) was added Pd/C (0.25 g, 10%) under N₂ atmosphere and then hydrogenated for 4h at RT. The reaction mixture was filtered through bed of celite and the filtrate was

concentrated under vacuum. The solid obtained was washed with 3% methanol in chloroform which gave the title product (4.1 g, 89%) as an off-white solid.

TLC: CHCl₃/MeOH, 4:1, R_F=0.1.

5 Example 7

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5'-O-Triisopropylsilyl-2'-deoxyuridine (7)

Imidazole (0.183 g, 2.69 mmol) was added to a solution of 2'-deoxyuridine (0.272 g, 1.19 mmol) in dry DMF (5 mL) under nitrogen. The mixture was cooled in an ice-salt bath before drop-wise addition of triisopropylsilyl chloride (0.28 mL, 1.31 mmol) via a syringe. The reaction mixture was kept at 0°C for 3h, allowed to warm up to room temperature andthen stirred at room temperature for 22h. After addition of water (5 mL), the crude mixture was extracted with CHCl₃ (2 x 10 mL). The organic layers were combined and dried over MgSO₄. Removal of the solvent under reduced pressure afforded a crude transparent oil which was further purified by flash chromatography eluting the column (ISOLUTE SI) a gradient of 0 \rightarrow 10% CH₃OH in CHCl₃. The title compound was obtained from the fractions with R_f = 0.25 (10% CH₃OH in CHCl₃) as a crystalline white solid (0.366 g, 74%). M.p. 152-153 °C.

¹H NMR (300 MHz, CDCl₃) δ 1.08 (21H, m, *i*Pr-H), 2.18 (1H, m, 2'-H), 2.49 (1H, m, 2'-H), 3.97 (2H, m, 5'-H), 4.08 (1H, m, 4'-H), 4.56 (1H, m, 3'-H), 5.70 (1H, d, J = 8.1 Hz, 5-H), 6.38 (1H, t, J = 6.2 Hz, 1'-H), 7.96 (1H, d, J = 8.1 Hz, 6-H), 10.16 (1H, bs, 3-NH). ¹³C NMR (75 MHz, CDCl₃) δ 12.7 (*i*Pr-CH), 18.4 (*i*Pr-CH₃), 41.9 (2'-CH₂), 63.8 (5'-CH₂), 71.7 (3'-CH), 85.7 (1'-CH), 88.0 (4'-CH), 102.5 (5-CH), 140.9 (6-CH), 151.1 (2-C), 164.5 (4-C).

ES⁺ m/z (%) 790 ([2M+Na]⁺, 10), 407 ([M+Na]⁺, 100). HRMS (ES⁺) Found [M+Na]⁺ 407.1988; C₁₈H₃₂N₂O₅SiNa⁺ requires 407.1973. Anal. Calcd for C₁₈H₃₂N₂O₅Si (%) C 56.22, H 8.39, N 7.28, found C 56.07, H 8.50, N 7.18.

Example 8

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5'-(4-pyridydiphenylmethyl)amino-2',5'-dideoxyuridine (8)

5'-Amino-2',5'-dideoxyuridine (0.108 g, 0.475 mmol) was added to a solution of diphenyl(4-pyridyl)chloromethane hydrochloride (0.150 g, 0.474 mmol), pyridine (3 mL) and Et₃N (0.12 mL, 0.862 mmol). The reaction mixture was heated at 40°C for 4h then the temperature was increased to 70°C for 10h. The crude solution was partitioned between water (5 mL) and EtOAc (3x7 mL). The organic extracts were combined, dried over Na₂SO₄ and concentrated in vacuo. The brown solid obtained was taken in MeOH and the remaining insoluble material was filtered off. The filtrate was concentrated under reduced pressure and further purified by flash column chromatography (ISOLUTE SI column) using a gradient elution of 0 \rightarrow 8% MeOH in CHCl₃. The fractions with R_f = 0.24 (10% MeOH/CHCl₃) afforded the title compound as a pale yellow solid (61 mg, 27%).

¹H NMR (300 MHz, CD₃OD) δ 2.15-2.60 (4H, m, 2',5'-H), 4.03 (1H, m, 3'-H or 4'-H), 4.18 (1H, m, 3'-H or 4'-H), 5.63 (1H, d, J = 8.0 Hz, 5-H), 6.21 (1H, t, J = 6.4 Hz, 1'-H), 7.20-7.48 (11H, m, 6-H and Ph-H), 7.59 (2H, d, J = 5.9 Hz, 4"-H), 8.42 (2H, d, J = 5.9 Hz, 5"-H). ¹³C NMR (75 MHz, CD₃OD) δ 40.9 (2'-CH₂), 47.6 (5'-CH₂), 72.2 (2"-C), 73.3 (3'-CH),

¹³C NMR (75 MHz, CD₃OD) δ 40.9 (2'-CH₂), 47.6 (5'-CH₂), 72.2 (2"-C), 73.3 (3'-CH), 87.1 (1'-CH or 4'-CH), 87.9 (1'-CH or 4'-CH), 103.4 (5-CH), 125.7 (4"-CH), 128.6 (Ph-CH), 129.7 (Ph-CH), 130.2 (Ph-CH), 130.3 (Ph-CH), 142.7 (6-CH), 145.8 (Ph-C), 146.1 (Ph-C), 150.2 (5"-CH), 152.4 (3"-C), 158.1 (2-C), 166.4 (4-C). ES⁺ m/z (%) 963 ([2M+Na]⁺, 13), 493 ([M+Na]⁺, 84), 471 ([M+H]⁺, 13). HRMS (ES⁺) Found [M+H]⁺ 471.2033; C₂₇H₂₇N₄O₄⁺ requires 471.2027.

M.p. 131-133°C.

Example 9

5'-O-trityl-2'-deoxyuridine (9)

2'-Deoxyuridine (1.00g, 4.39 mmol) and triphenylmethylchloride (1.34 g, 4.83mmol) were stirred in dry pyridine (20 ml) overnight at 50°C under an atmosphere of nitrogen. The reaction mixture was then poured into ice-H₂O (100 ml) with vigorous stirring and filtered. The precipitate was dissolved in EtOAc (100 ml) and the solution was washed with 0.5M HCl (100ml) and H₂O (100 ml), dried (Na₂SO₄) and reduced *in vacuo*. The residue was washed with toluene to leave the title compound (1.99 g, 97%) as a pale yellow solid. For analytical purposes, the compound was purified by gradient flash column chromatography, eluting with 5→10% MeOH/CHCl₃.

¹H NMR (300MHz;CDCl₃): δ 2.34 (1H, m, 2'-H), 2.45 (1H, m, 2'-H), 3.51 (2H, ddd, *J* = 3.5, 8.6, 10.6 Hz, 5'-H), 4.12 (1H, dd, *J* = 3.6, 7.2 Hz, 4'-H), 4.64 (1H, m, 3'-H), 5.47 (1H, d, *J* = 8.1 Hz, 5-H), 6.40 (1H, t, *J* = 6.3 Hz, 1'H), 7.22-7.49 (15H, m, Ph-H), 7.86 (1H, d, *J* = 8.1Hz, 6-H), 9.37 (1H, s, 3-NH); ¹³C NMR (75MHz;CDCl₃): δ 41.60 (2'-CH₂), 63.53 (5'-CH₂), 71.84 (3'-CH), 85.49 (4'-CH), 86.43 (1'-CH), 88.03 (Ph₃C), 127.92 (Ph-CH), 128.68 (Ph-CH), 129.49 (Ph-CH), 140.69 (6-CH), 143.67 (Ph-CH), 153.24 (2-C), 163.93 (4-C). MS (AP⁺., *m/z*): 243 (Tr⁺, 100%); R_f (10% MeOH/CHCl₃): 0.49;

Example 10

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1-[4-(*tert*-Butyl-dimethyl-silanyloxy)-5-trityloxymethyl-tetrahydro-furan-2-yl]-1*H* pyrimidine-2,4-dione or 3'-O-*tert*-Butylsilyl-5'-O-trityl-2'-deoxyuridine (10) 5'-O-trityl-2'deoxyuridine (0.70 g, 1.49 mmol) in dry DMF (3 ml) was added drop-wise under an atmosphere of nitrogen, with ice bath cooling, to a stirred solution of *tert*-butyldimethylsilyl chloride (0.25 g, 1.65 mmol) and imidazole (0.22 g, 3.28 mmol) in dry DMF (3 ml). The mixture was allowed to warm to room temperature and stirred ovemight. H₂O (10 ml) was added (10ml) and the mixture was extracted with Et₂O (2x50 ml). The combined extracts were washed with saturated NaHCO₃ (50 ml) and H₂O (50 ml), dried (Na₂SO₄) and reduced *in vacuo*. A flash silica column eluting with 3% MeOH/CHCl₃ gave the title compound (0.65 g, 74%) as white foam.

¹H NMR (300MHz;CDCl₃): δ -0.05 [3H, s, Si(CH₃)₂] and 0.00 [3H, s, Si(CH₃)₂], 0.85 [9H, s, C(CH₃)₃], 2.12-2.20 (1H, m, 2'-H), 2.31-2.39 (1H, m, 2'-H), 3.33 (1H, dd, J = 2.8, 10.7 Hz, 5'-H), 3.46 (1H, dd, J = 2.9, 10.7Hz, 5'-H), 3.92 (1H, dt, J = 2.8, 4.4Hz, 4'-H), 4.51 (1H, dd, J = 4.9, 10.9Hz, 3'-H), 5.34 (1H, d, J = 8.1Hz, 5-H), 6.26 (1H, t, J = 6.0Hz, 1'-H), 7.23-7.39 (15H, m, Ph-H), 7.85 (1H, d, J = 8.1Hz, 6-H), 9.11 (1H, s, 3-NH); ¹³C NMR (75MHz; CDCl₃): -4.49 and -4.20 (Si(CH₃)₂), 18.37[C(CH₃)₃], 22.06 and 26.17 [C(CH₃)₃], 42.23 (2'-CH₂), 62.27 (5'-CH₂), 71.38(3'-CH), 85.55 (4'-CH) 86.83 (1'-CH), 87.89 (Ph₃C), 102.70 (5-CH), 127.91 (Ph-CH), 128.48 (Ph-CH), 129.15 (Ph-CH), 140.62 (6-CH), 143.59 (Ph-CH), 150.68 (2-C), 163.81 (4-C); MS (AP⁺., m/z): 243 (Tr⁺, 50%), 341 (M-Tr⁺, 75%), 607 (M+Na⁺, 100%) R_ℓ (3%MeOH/CHCl₃): 0.33;

Example 11

5'-O-Triphenylsilyl-2'-deoxyuridine (11)

A solution of triphenylsilyl chloride (0.437 g, 1.48 mmol) in dry pyridine (4 mL) was added drop-wise to a solution of 2'-deoxyuridine (0.278 g, 1.22 mmol) in dry pyridine (4 mL) previously cooled in an ice-salt bath. The reaction mixture was kept at 0°C for 1h.

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The reaction was monitored by TLC (10% CH₃OH in CHCl₃) and quenched with CH₃OH (50 L). The solvent was removed under reduced pressure to give a crude yellow liquid which was further purified by silica gel column chromatography (Isolute SI column) using a gradient elution of $0 \rightarrow 10\%$ CH₃OH in CHCl₃. The fractions with R_f = 0.30 (10% CH₃OH/CHCl₃) were combined and concentrated to yield the title compound as a white crystalline solid (0.506 g, 85%).

¹H NMR (300MHz, CDCl₃) δ 2.25 (1H, m, 2'-H), 2.44 (1H, m, 2'-H), 2.95 (1H, bs, 3'-OH), 3.93-4.27 (3H, m, 5'-H and 4'-H), 4.60 (1H, m, 3'-H), 5.19 (1H, d, J = 8.2 Hz, 5-H), 6.41 (1H, t, J = 6.4 Hz, 1'-H), 7.35-7.73 (15H, m, Ph-H), 7.80 (1H, d, J = 8.1 Hz, 6-H), 9.46 (1H, bs, 3-NH).

¹³C NMR (75MHz, CDCl₃) δ 41.6 (2'-CH₂), 63.8 (5'-CH₂), 71.7 (3'-CH), 85.3 (1'-CH), 87.3 (4'-CH), 102.7 (5-CH), 128.6 (Ph-CH), 131.1 (Ph-CH), 133.3 (Ph-C), 135.8 (Ph-CH), 140.5 (6-CH), 150.9 (2-C), 163.9 (4-C).

15 ES⁺ m/z (%) 509 ([M+Na]⁺, 100).

ES⁺ m/z (%) 509 ([M+Na]⁺, 78), 151 (100).

HRMS (ES⁺) Found [M+Na]⁺ 509.1504; C₂₇H₂₆N₂O₅Si requires 509.1503.

Anal. calcd for C₂₇H₂₆N₂O₅Si (%): 0.32 HCl C 65.09, H 5.32, N 5.62 found: C 65.01, H 5.27, N 5.62.

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Example 12

5'-O-tert-Butyldiphenylsilyl-2'-deoxyuridine (12)

2'-Deoxyuridine (0.530 g, 2.32 mmol) was dissolved in dry DMF (5mL) under nitrogen and the solution was cooled in an ice-salt bath. A solution of *tert*-butyldiphenylsilyl chloride (0.710 g, 2.58 mmol) and imidazole (0.342 g, 5.69 mmol) in dry DMF (4 mL) was then added drop-wise. The reaction mixture was stirred at 0°C for 2h and then at room temperature for 15h. The reaction was quenched by addition of water (15 mL).

The crude mixture was extracted with CHCl₃ (2 x 15 mL). The organic layers were combined, dried over MgSO₄ and concentrated *in vacuo* to give a transparent oil (0.419 g). This oil was chromatographed on a silica gel column (Isolute SI column) eluted with a gradient of $0 \rightarrow 10\%$ CH₃OH in CHCl₃. The fractions with R_f = 0.26 (10%

5 CH₃OH/CHCl₃) were gathered and concentrated to afford the title compound as a white crystalline solid (0.823 g, 76%).

¹H NMR (300MHz, CDCl₃) δ 1.14 (9H, m, tBu-H), 2.27 (1H, m, 2'-H), 2.50 (1H, m, 2'-H), 2.69 (1H, bs, 3'-OH), 3.90 (1H, m, 4'-H), 4.05 (2H, m, 5'-H), 4.60 (1H, m, 3'-H), 5.52 (1H, d, J = 8.1 Hz, 5-H), 6.41 (1H, t, J = 6.4 Hz, 1'-H), 7.48 (6H, m, Ph-H), 7.70 (4H, m, Ph-H), 7.87 (1H, d, J = 8.1 Hz, 6-H), 9.34 (1H, bs, 3-NH).

¹³C NMR (75MHz, CDCl₃) δ 19.7 (*t*Bu-C), 27.4 (*t*Bu-CH₃), 41.7 (2'-CH₂), 64.1 (5'-CH₂), 71.9 (3'-CH), 85.4 (1'-CH), 87.5 (4'-CH), 102.6 (5-CH), 128.4 (Ph-CH), 128.5 (Ph-CH), 130.6 (Ph-CH), 132.7 (Ph-C), 133.1 (Ph-C), 135.8 (Ph-CH), 136.0 (Ph-CH), 140.5 (6-CH), 150.9 (2-C), 163.9 (4-C).

15 ES⁺ m/z (%) 489 ([M+Na]⁺, 100).

Anal. calcd for C₂₅H₃₀N₂O₅Si 0.58 (%):HCl C 61.56, H 6.32, N 5.74, found C 61.61, H 6.23, N 5.72.

Example 13

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3',5'-O-bistertbutyldimethylsilyl-2'-deoxyuridine

A solution of *t*-butyl dimethylsilylchloride (2.18 g, 14.46 mmol) and imidazole (1.07 g, 28.92 mmol) in DMF (30 ml) was added slowly (drop wise) to a stirred solution of 2'-deoxyuridine (3 g, 13.15 mmol) in dry DMF (40 ml), with ice-bath cooling at 0°C, under atmosphere of nitrogen.

After 2 hours, H₂O (100 ml) was added and the mixture was extracted with AcOEt (3×100 ml). The combined extracts were washed with saturated NaHCO₃ (2×100 ml), dried (MgSO₄) and concentrated. The residue was purified by flash chromatography

and the title compound was isolated as a white amorphous solid from the fractions with Rf= 0.65 (10% CH₃OH in CHCl₃).

¹H-NMR (300MHz, CDCl₃) δ 0.2 (s, 12H, tBu[C*H*₃]₂Si), 1.0 (s, 18H, *tBu*[CH₃]₂Si), 2.5 (1H, m, 2'-H), 2.2 (1H, m, 2'-H), 3.06 (1H, d, *J*=5.0 Hz, 3'-H), 3.95 (1H, dd, *J*=11.5 Hz, 2.2 Hz, 5'-H), 4.01 (1H, dd, *J*=11.5, 2.6 Hz, 5'-H), 4.15 (1H, m, 4'-H), 5.78 (1H, d, *J*=8.23 Hz, 5-H), 6.45 (1H, t, *J*=6.95 Hz, 1'-H), 8.02 (1H, d, *J*=8.2 Hz, 6-H), 9.5 (1H, s, 3-NH).

¹³C-NMR (75MHz, CDCl₃) δ 163.9 (4-C), 150.9 (2-C), 140.8 (6-CH), 102.7 (5-CH), 87.9 (4'-CH), 85.8 (1'-CH), 72.5 (3'-CH), 63.7 (5'-CH₂), 42.0 (2'-CH₂), 26.3 (CH₃), 18.8 (CH₃).

Example 14

5'-Tritylamino-2',5'-dideoxyuridine (14)

5'-Amino-2',5'-dideoxyuridine (0.200 g, 0.88 mmol) was taken in dry pyridine (5 mL) and the mixture was sonicated for a few minutes. Trityl chloride (0.278 g, 1.00 mmol) was added and the reaction mixture was stirred at 50 °C overnight. The reaction was then quenched with water (20 mL). The crude mixture was extracted with DCM (3 x 10 mL). The organic layers were combined, washed with water (10 mL), dried over MgSO₄ and concentrated on the rotary evaporator. The resultant brown oil was further purified by silica gel column chromatography (Isolute SI column) using a gradient elution of 0 → 10% CH₃OH in CHCl₃. The fractions with R_f = 0.28 (10% CH₃OH/CHCl₃) were pooled and evaporated to dryness to yield the title compound as a white solid (0.202 g, 49%).

¹H NMR (300MHz, CDCl₃) δ 2.07 (2H, m, 2'-H), 2.28-2.53 (2H, m, 5'-H), 2.73 (1H, dd, J = 3.5, 12.1 Hz, 1"-NH), 2.97 (1H, bs, 3'-OH), 4.19 (1H, m, 4'-H), 4.33 (1H, m, 3'-H), 5.72 (1H, d, J = 8.1 Hz, 5-H), 6.36 (1H, t, J = 6.4 Hz, 1'-H), 7.14 (1H, d, J = 8.1 Hz, 6-H), 7.23-7.43 (9H, m, Ph-H), 7.57 (6H, m, Ph-CH), 9.47 (1H, bs, 3-NH).

¹³C NMR (75MHz, CDCl₃) δ 40.8 (2'-CH₂), 46.6 (5'-CH₂), 71.1 (2"-C), 73.0 (3'-CH), 85.4 (1'-CH), 86.7 (4'-CH), 103.2 (5-CH), 127.0 (Ph-CH), 128.4 (Ph-CH), 129.0 (Ph-CH), 139.8 (6-CH), 145.8 (Ph-C), 150.7 (2-C), 163.7 (4-C).

ES⁺ m/z (%) 243 (Ph₃C⁺, 100), 470 ([M+H]⁺, 4), 492 ([M+Na]⁺, 23).

5 HRMS (ES⁺) Found [M+H]⁺ 470.2076; C₂₈H₂₈N₃O₄ requires 470.2074. M.p. 132-134°C.

Anal calcd for C₂₈H₂₇N₃O₄ (%): 0.53 HCl C 68.79, H 5.68, N 8.60 found: C 68.79, H 5.55, N 8.59.

10 <u>Example 15</u>

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3'-O-tertbutyldimethylsilyl-5'-Tritylamino-2',5'-dideoxyuridine (15)

A solution of 5'-tritylamino-2',5'-dideoxyuridine (0.172 g, 0.37 mmol) in anhydrous DMF (2 mL) was added drop-wise to an ice cold solution of *tert*-butyl dimethylsilyl chloride (68 mg, 0.45 mmol) and imidazole (60 mg, 0.88 mmol) in anhydrous DMF (2 mL). The reaction mixture was stirred at 0°C for 2h and at room temperature for a further 20h. It was then partitioned between water (10 mL) and Et₂O (2 x 20 mL). The combined organic layers were washed with a saturated aqueous solution of NaHCO₃ (15 mL), dried over MgSO₄ and concentrated *in vacuo*. The resultant white solid was further purified by column chromatography (Isolute SI column) using a gradient elution of 0 \rightarrow 10% CH₃OH in CHCl₃. The fractions with R_f = 0.69 (10% CH₃OH/CHCl₃) were pooled and evaporated to dryness to yield the title compound as a white solid (154 mg, 72%).

¹H NMR (300MHz, CDCl₃) δ 0.00-0.02 (6H, 2 x s, Si(CH₃)₂), 0.86 (9H, s, C(CH₃)₃), 1.90 (2H, m, 2'-H), 2.11-2.35 (2H, m, 5'-H), 2.59 (1H, bd, $J \approx 13$ Hz, 1"-NH), 4.06 (2H, m, 3'-H) and 4'-H), 5.65 (1H, d, J = 8.1 Hz, 5-H), 6.25 (1H, t, J = 6.3 Hz, 1'-H), 7.07 (1H, d, J = 8.1 Hz, 6-H), 7.14-7.37 (9H, m,Ph-H), 7.48 (6H, m,).

¹³C NMR (75MHz, CDCl₃) δ -4.4 (Si<u>C</u>H₃), -4.2 (Si<u>C</u>H₃), 18.4 (<u>C</u>(CH₃)₃), 26.1 (C(<u>C</u>H₃)₃), 41.4 (2'-CH₂), 46.5 (5'-CH₂), 71.2 (2"-C), 73.3 (3'-CH), 85.6 (1'-CH), 87.3 (4'-CH), 103.1 (5-CH), 127.0 (Ph-CH), 128.4 (Ph-CH), 129.0 (Ph-CH), 139.7 (6-CH), 145.9 (Ph-C), 150.6 (2-C), 163.7 (4-C).

5 ES⁺ m/z (%) 584 ([M+H]⁺), 606 ([M+Na]⁺).

HRMS (ES⁺) Found [M+H]⁺ 584.2938; C₃₄H₄₂N₃O₄Si requires 584.2939.

Example 16

1-(4-Fluoro-5-trityloxymethyl-tetrahydro-furan-2-yl)2,3-dihydro-1*H*-pyrimidin-4-one or 3'-Fluoro-5'-O-trityl-2',3'-dideoxyuridine (16)

3'-Fluoro-2',3'-dideoxyuridine (0.3 g, 1.30 mmol) and triphenylmethyl chloride (0.44 g, 1.57 mmol) were stirred in dry pyridine (20 ml) overnight at 50°C under an atmosphere of nitrogen. The reaction mixture was then poured into ice- H_2O (50 ml) with vigorous stirring and filtered. The precipitate was dissolved in EtOAc (50 ml) and the solution was washed with 0.5M HCl (50 ml) and H_2O (50 ml) dried (Na₂SO₄) and reduced *in vacuo* to obtain a crude product, which was purified by gradient flash column chromatography eluting with $2\rightarrow6\%$ MeOH/CHCl₃ to obtain the title compound as a white solid (0.48 g, 77%).

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¹H NMR(300MHz;CDCl₃): δ 2.27-2.50 (1H, m, 2'-H), 2.78-2.92 (1H, m, 2'-H), 3.53-3.63 (2H, m, 5'H), 4.41-4.51 (1H, d, J = 27.3 Hz, 4'-H), 5.33-5.53 (2H, m,3',5-H), 6.50-6.55 (1H, m, 1'-H), 7.46 (15H, m, Ph-H), 7.80 (1H, d, J 8.1, 6-H);

¹³C NMR(75MHz;CDCl₃): δ 39.43 and 39.71 (2'-CH₂), 63.75 and 63.89 (5'-CH₂), 84.54 25 and 84.88 (4'-CH), 85.38 (1'-CH), 88.27 (Ph-C) -, 93.44 (Ph-CH), 95.80 (Ph-CH), J 178.48, 3'-CH), 103.08 (5-CH), 128.06 (Ph-CH), 128.58 (Ph-CH), 129.00 (Ph-CH), 140.18 (6-CH), 143.31 (Ph-C), 150.67 (2-C), 163.53 (4-C);

¹⁹F NMR(282MHz;CDCl₃):δ -174.26;

MS (CI/NH₃., m/z): 473.2 (M+H⁺, 50%), 490.3 (M+NH₄⁺, 80%);

30 HRMS (El., M⁺): Calculated for C₂₈H₂₅N₂O₄F, requires 472.1798, found 472.1797.

IR_{max}/cm⁻¹ (KBr): 703 (s) and 763(s) (Aromatic, monosubstituted), 1107.9 (C-F), 1689.3 (C=O) and 1702.3 (C=O).R_f (10% MeOH / CHCl₃): 0.52. Mp: 128-130°C.

5 Example 17

3'-Fluoro-5'-tritylamino-2',3',5'-trideoxyuridine (17)

The title compound was obtained as a light yellow crystalline solid (91 mg, 32%) from the reaction of the corresponding amine (0.137 g, 0.59 mmol) and trityl chloride (0.199 g, 0.66 mmol) in dry pyridine (4mL). The procedure was similar to that followed for the preparation of the 3' hydroxy analogue 5'tritylamino-2',5'-dideoxyuridine (WSP871, see example 14).

¹*H NMR* (300MHz, CDCl₃) δ 1.87-2.13 (2H, m, 2'-H), 2.28 (1H, dd, J = 8.1, 12.0 Hz, 1"-NH), 2.57-2.78 (2H, m, 5'-H), 4.48 (1H, dm, J ≈ 25 Hz, 4'-H), 5.11 (1H, dd, J = 5.3, 53.7 Hz, 3'-H), 5.71 (1H, d, J = 8.1 Hz, 5-H), 6.37 (1H, dd, J = 5.6, 8.7 Hz, 1'-H), 6.98 (1H, d, J = 8.1 Hz, 6-H), 7.23-7.43 (9H, m, Ph-H), 7.53 (6H, m, Ph-CH), 9.39 (1H, s, 3-NH). ¹³C *NMR* (75MHz, CDCl₃) δ 38.5 (d, J = 21.8 Hz, 2'-CH₂), 46.1 (d, J = 9.2 Hz, 5'-CH₂), 71.1 (Ph₃C), 85.2 (d, J = 25.3 Hz, 4'-CH), 85.5 (1'-CH), 94.4 (d, J = 179.9 Hz, 3'-CH), 103.6 (5-CH), 127.1 (Ph-CH), 128.5 (Ph-CH), 128.9 (Ph-CH), 139.3 (6-CH), 145.7 (Ph-C), 150.5 (2-C), 163.4 (4-C).

¹⁹F NMR (282MHz, CDCl₃) δ -175.7 (m, 3'-F). ES⁺ m/z (%) 243 (Ph₃C⁺, 93), 494 ([M+Na]⁺, 92).

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Example 18

3'-Fluoro-5'-O-triphenylsilyi-2',3'-dideoxyuridine(18)

The title compound was synthesised following a similar procedure as described for Example 11. 3'-Fluoro-2',3'-dideoxyuridine (0.214 g, 0.93 mmol) was reacted with triphenylsilyl chloride (0.332 g, 1.12 mmol) in dry pyridine (7 mL) for 3h. to yield the title compound as a white solid (0.274 g, 60%).

¹H NMR (300MHz, CDCl₃) δ 2.19 (1H, m, 2'-H), 2.67 (1H, m, 2'-H), 4.11 (2H, m, 5'-OH), 4.36 (1H, d, J = 27.1 Hz, 3'-H), 5.22 (1.5H, m, 4'-H and 5-H), 5.40 (0.5H, d, J = 4.8 Hz, 4'-H), 6.50 (1H, dd, J = 5.4, 9.1 Hz, 1'-H), 7.41-7.75 (16H, m, 6-H and Ph-H), 9.04 (1H, bs, 3-NH).

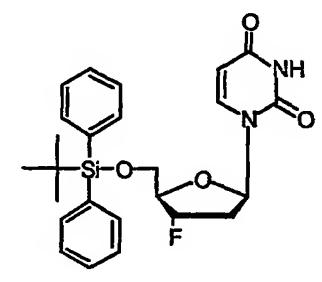
¹³C NMR (75MHz, CDCl₃) δ 39.5 (d, J = 20.7 Hz, 2'-CH₂), 64.3 (d, J = 11.5 Hz, 5'-CH₂), 85.2 (1'-CH), 85.4 (d, J = 24.7 Hz, 4'-CH), 94.9 (d, J = 178.7 Hz, 3'-CH), 103.1 (5-CH), 128.8 (Ph-CH), 131.2 (Ph-CH), 133.0 (Ph-C), 135.7 (Ph-CH), 140.1 (6-CH), 150.7 (2-C), 163.5 (4-C).

¹⁹F NMR (282MHz, CDCl₃) δ -175.1 (m, 3'-F). ES⁺ m/z (%) 511 ([M+Na]⁺, 5), 87 (100). ES⁻ m/z (%) 487 ([M-H⁺], 31), 75 (100).

20 <u>Example 19</u>

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3'-Fluoro-5'-O-tert-Butyldiphenylsilyl-2',3'-dideoxyuridine (19)

The title compound was synthesised following a similar procedure as described for Example 12. 3'-Fluoro-2',3'-dideoxyuridine (0.176 g, 0.77 mmol) was reacted with tert-

butyldiphenylsilyl chloride (0.238 g, 0.87 mmol) and imidazole (0.116 g, 1.70 mmol) in dry DMF (4 mL) for 3h. Compound WSP948 was obtained as a white solid (0.331 g, 92%).

¹*H NMR* (300MHz, CDCl₃) δ 1.17 (9H, m, tBu-H), 2.24 (1H, m, 2'-H), 2.78 (1H, m, 2'-H), 4.00 (2H, m, 5'-H), 4.38 (1H, d, J = 26.7 Hz, 4'-H), 5.34 (1H, dd, J = 4.9, 53.8 Hz, 3'-H), 5.56 (1H, d, J = 8.1 Hz, 5-H), 6.51 (1H, m, 1'-H), 7.43-7.60 (6H, m, Ph-H), 7.65-7.74 (4H, m, Ph-H), 7.27 (1H, d, J = 8.1 Hz, 6-H), 9.11 (1H, bs, 3-NH). ¹³*C NMR* (75MHz, CDCl₃) δ 19.7 (tBu-C), 27.4 (tBu-CH₃), 39.7 (d, J = 21.3 Hz, 2'-CH₂), 64.1 (d, J = 10.9 Hz, 5'-CH₂), 85.4 (1'-CH), 85.6 (d, J = 24.7 Hz, 4'-CH), 94.7 (d, J = 178.7 Hz, 3'-CH), 103.2 (5-CH), 128.5 (Ph-CH), 128.6 (Ph-CH), 130.7 (Ph-CH), 132.2 (Ph-C), 132.8 (Ph-C), 135.7 (Ph-CH), 136.0 (Ph-CH), 140.0 (6-CH), 150.6 (2-C), 163.5 (4-C).

¹⁹F NMR (282MHz, CDCl₃) δ -175.5 (m, 3'-F). ES m/z (%)467 ([M-H⁺], 53), 75 (100).

Example 20

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5'-O-paramethoxytrityl-2'-deoxyuridine (20)

4-Methoxytrityl (0.610g, 1.98 mmol) was added to a solution of 2'-deoxyuridine (0.410g, 1.80 mmol) in anhydrous pyridine (10 mL). The reaction mixture was stirred at 50°C for 40h. The crude mixture was partitioned between water (40 mL) and DCM (2 x 40 mL). The organic layers were combined, washed with water (2 x 80 mL), dried over MgSO₄ and concentrated *in vacuo*. The resultant yellow oil was further purified by silica gel column chromatography (using Jones Chromatography Isolute SI columns). The column was eluted with a gradient of $0 \rightarrow 5\%$ CH₃OH in CHCl₃. The fractions with R_f = 0.28 (10% CH₃OH/CHCl₃) yielded the title compound as a white crystalline solid (0.625 g, 69%).

¹H NMR (300MHz, CDCl₃) δ 2.27 (1H, m, 5'-H), 2.42 (1H, m, 5'-H), 2.57 (1H, bs, 3'-OH), 3.42 (2H, m, 2'-H), 3.76 (3H, s, OCH₃), 4.00 (1H, m, 4'-H), 4.54 (1H, m, 3'-H), 5.37 (1H,

d, J = 8.1 Hz, 5-H), 6.29 (1H, t, J = 6.3 Hz, 1'-H), 6.82 (2H, m, Ar-H), 7.18-7.38 (12H, m, Ar-H), 7.74 (1H, d, J = 8.1 Hz, 6-H), 9.20 (1H, bs, 3-NH).

¹³C NMR (75MHz, CDCl₃) δ 41.6 (2'-CH₂), 55.7 (OCH₃), 63.5 (5'-CH₂), 71.9 (3'-CH), 85.5 (4'-CH), 86.5 (1'-CH), 87.8 (Ar₃C), 102.7 (5-CH), 113.8 (Ar₂-CH), 127.7 (Ar₂-CH), 128.5 (Ar₂-CH), 128.8 (Ar₂-CH), 130.8 (Ar₂-CH), 135.1 (Ar₂-C), 140.6 (6-CH), 144.1 (Ar₂-C), 144.5 (Ar₂-C), 140.6 (6-CH), 144.1 (Ar₂-C), 140.6 (6-CH), 140.6 (6-

144.3 (Ar-C), 150.8 (2-C), 159.3 (Ar-C), 163.7 (4-C).

ES⁺ m/z (%) 523 ([M+Na]⁺, 100)

HRMS (ES⁺) Found [M+Na]⁺ 523.1848; C₂₉H₂₈N₂O₆Na requires 523.1845.

IR (KBr) 3208, 3054, 1714, 1694, 1682, 1507, 1470, 1250, 1092, 1035, 759 cm⁻¹.

10 M.p. 96-97°C.

5

Anal calcd for $C_{29}H_{28}N_2O_6$ (%): 1.43 HCl, 0.40 H_2O C 62.21, H 5.44, N 5.00, Cl 9.06; found: C 62.17, H 5.05, N 4.85, Cl 8.86.

Example 21

NC NO NO OH

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5'-O-(4-cyanotrityl)-2'-deoxyuridine (21)

4-Cyanotrityl (0.397 g, 1.31 mmol) was added to a solution of 2'-deoxyuridine (0.229 g, 1.00 mmol) in dry pyridine (5 mL). As the reaction was not complete after 72h at 50°C, DMAP (11 mg, 0.09 mmol) was added and the reaction mixture was kept at 50°C for a further 20h. H_2O (20 mL) was added and the crude mixture was extracted with DCM (2 x 15 mL and 10 mL). The combined organic layers were dried over Na_2SO_4 , concentrated *in vacuo* and purified by flash column chromatography eluting the column (ISOLUTE SI) with a gradient of $O \rightarrow 6\%$ CH₃OH in CHCl₃. The fractions with $R_f = 0.29$ (10% CH₃OH/CHCl₃) afforded the title compound as a white solid (0.215 g, 43%).

25

¹H NMR (300 MHz, CDCl₃) δ 2.19 (1H, m, 2'-C<u>H</u>H), 2.45 (1H, m, 2'- CH<u>H</u>), 2.94 (1H, bs, 3'-OH), 3.38 (2H, m, 5'-H), 4.06 (1H, m, 4'-H), 4.50 (1H, m, 3'-H), 5.45 (1H, d, J = 8.1 Hz, 5-H), 6.27 (1H, t, J = 6.2 Hz, 1'-H), 7.24-7.34 (10H, m, Ph-H), 7.53-7.60 (5H, m, 6-H and Ar-H), 9.50 (1H, bs, 3-NH).

¹³C NMR (75 MHz, CDCl₃) δ 41.4 (2'-CH₂), 63.9 (5'-CH₂), 71.7 (3'-CH), 85.6 (1'-CH), 86.1 (4'-CH), 87.6 (ArPh₂C), 102.8 (5-CH), 111.4 (Ar-C), 119.0 (C \equiv N), 128.6 (Ph-CH), 128.8 (Ph-CH), 129.0 (Ar-CH), 132.4 (Ar-CH), 140.3 (6-CH), 142.0 (Ph-C), 142.1 (Ph-C), 150.1 (Ar-C), 150.8 (2-C), 163.8 (4-C).

5 ES⁺ m/z (%) 518 ([M+Na]⁺, 23), 268 (CNTr⁺, 100). ES⁻ m/z (%) 494 (M-H⁺, 100). HRMS (ES⁺) Found [M+NH₄]⁺ 513.2132; C₂₉H₂₉N₄O₅⁺ requires 513.2132. M.p. 92-95°C.

IR (KBr) 3401, 3180, 3060, 2230 (CN), 1685, 1463, 1273, 1088 cm⁻¹.

10 Anal calcd for C₂₉H₂₅N₃O₅ (%): 2.35 HCl C 59.93, H 4.74, N 7.23 found: C 59.89, H 4.45, N 7.02.

Example 22

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15 5'-[(4-cyanotrityl)amino]-2',5'-dideoxyuridine (22)

4-Cyanotrityl chloride (0.406 g, 1.34 mmol) was added to a solution of 5'-amino-2',5'-dideoxyuridine (0.239 g, 1.05 mmol) in dry pyridine (5 mL). The reaction mixture was stirred at 40°C for 48h. The reaction mixture was filtered and the filtrate concentrated *in vacuo*. Purification was carried out using flash column chromatography eluting the column (ISOLUTE SI) with a gradient of $0 \rightarrow 5\%$ CH₃OH in CHCl₃. The fractions with R_f = 0.31 (10% CH₃OH/CHCl₃) afforded the title compound as a white crystalline solid (0.386 g, 37%).

¹H NMR (300 MHz, CDCl₃) δ 2.03 (2H, m, 2'-C<u>H</u>H and 5'-N<u>H</u>), 2.21 (1H, m, 5'-C<u>H</u>H), 2.37 (1H, m, 2'-C<u>H</u>H),2.58 (1H, m, 5'-C<u>H</u>H), 3.31 (1H, bs, 3'-OH), 4.10 (1H, m, 4'-H), 4.24 (1H, m, 3'-H), 5.63 (1H, d, J = 8.1 Hz, 5-H), 6.24 (1H, t, J = 6.3 Hz, 1'-H), 7.00 (1H, dd, J = 2.1, 8.1 Hz, 6-H), 7.18-7.48 (10H, m, Ph-H), 7.54 (2H, d, J = 8.1 Hz, Ar-H), 7.66 (2H, d, J = 8.1 Hz, Ar-H), 9.74 (1H, bs, 3-NH).

¹³C NMR (75 MHz, CDCl₃) δ 40.7 (2'-CH₂), 46.5 (5'-CH₂), 71.3 (A_fPh₂C), 72.8 (3'-CH), 85.6 (1'-CH), 86.5 (4'-CH), 103.3 (5-CH), 110.7 (A_f-CH), 119.2 (<u>C</u>N), 127.6 (Ph-CH), 128.7 (Ph-CH), 128.9 (Ph-CH), 129.1 (Ph-CH), 129.5 (A_f-CH), 132.4 (A_f-CH), 139.9 (6-CH), 144.3 (Ph-C), 145.0 (Ph-C), 150.8 (2-C), 151.6 (A_f-C), 163.8 (4-C).

5 ES⁺ m/z (%) 517 ([M+Na]⁺, 19), 495 ([M+H]⁺, 9), 517 (CNTr⁺, 100).
ES⁻ m/z (%) 493 (M-H⁺, 19), 111 (uracil-H⁺, 100).
HRMS (ES⁺) Found [M+H]⁺ 495.2023; C₂₉H₂₇N₄O₄⁺ requires 495.2027.
M.p. 160-163°C.

IR (KBr) 3387, 3177, 3027, 2230 (CN), 1699, 1661, 1466, 1267, 1097, 1039 cm⁻¹.

10 Anal calcd for C₂₉H₂₉N₃O₅, 0.98 (%): HCl, 0.40 H₂O C 64.80, H 5.21, N 10.42, Cl 6.46; found: C 64.69, H 4.90, N 10.28, Cl 6.84.

Example 23

15 5'-paraMethoxytritylamino-2',5'-dideoxyuridine (23)

The procedure was similar to that described for example 20. 5'-Amino-2',5'-dideoxyuridine (0.204 g, 0.90 mmol) was reacted with 4-methoxytrityl (0.292 g, 0.99 mmol) to yield the title compound as a white solid (0.115g, 26%).

¹H NMR (300MHz, CDCl₃) δ 2.04 (2H, m, 2'-H), 2.29-2.48 (2H, m, 5'-H), 2.69 (1H, dd, J = 3.7, 12.1 Hz, 5'-NH), 3.83 (3H, s, OCH₃), 4.15 (1H, m, 4'-H), 4.30 (1H, m, 3'-H), 5.69 (1H, d, J = 8.1 Hz, 5-H), 6.32 (1H, t, J = 6.4 Hz, 1'-H), 6.88 (2H, m, Ar-H), 7.17 (1H, d, J = 8.1 Hz, 6-H), 7.21-7.61 (12H, m, Ar-H).

 13 C NMR (75MHz, CDCl₃) δ 40.8 (2'-CH₂), 46.6 (5'-CH₂), 55.7 (OCH₃), 70.7 (Ar₃C), 73.0 (3'-CH), 85.4 (4'-CH), 86.8 (1'-CH), 103.2 (5-CH), 113.7 (Ar-CH), 126.9 (Ar-CH), 128.4 (Ar-CH), 128.9 (Ar-CH), 130.2 (Ar-CH), 138.0 (Ar-C), 139.9 (6-CH), 146.2 (Ar-C), 150.8

(2-C), 158.4 (Ar-C), 163.8 (4-C).

25

ES⁺ m/z (%) 522 ([M+Na]⁺, 27).

HRMS (ES⁺) Found [M+H]⁺ 500.2174; C₂₉H₃₀N₃O₅⁺ requires 500.2180. IR (KBr) 3052, 1713, 1694, 1682, 1666, 1650, 1506, 1250, 1034, 760 cm⁻¹.

M.p. 140-142°C.

TLC (10% $CH_3OH/CHCl_3$) $R_f = 0.29$.

Anal (%) found C 65.69, H 5.52, N 7.86, Cl 6.05;

Calcd for C₂₉H₂₉N₃O₅, 0.87 HCl C 65.56, H 5.67, N 7.91, Cl 5.81.

5

15

Example 24

5'-[(2-chlorotrityl)amino]-2',5'-dideoxyuridine (24)

5'-Amino-2',5'-dideoxyuridine (0.237 g, 1.04 mmol) was reacted with 2-chlorotrityl 10 chloride (0.415 g, 1.33 mmol) in dry pyridine (5 mL) at 40°C for 24h. A second portion of 2-chlorotrityl chloride (0.198 g, 0.63 mmol) was added. After a further 2h stiring at 40°C, the reaction was quenched with MeOH (2 mL). The reaction mixture was concentrated in vacuo and purified by flash column chromatography using an ISOLUTE SI column eluted with a gradient of $0 \rightarrow 5\%$ CH₃OH in CHCl₃. The fractions with R_f = 0.17 (10%) CH₃OH/CHCl₃) yielded the title compound as a white solid (85 mg, 16%).

¹H NMR (300 MHz, CDCl₃) δ 2.02 (1H, m, 2'-H), 2.20 (1H, m, 5'-H), 2.37 (1H, m, 2'-H), 2.53 (1H, m, 5'-H), 4.16 (1H, m, 3'-H), 4.24 (1H, m, 4'-H), 5.62 (1H, d, J = 8.1 Hz, 5-H), 6.26 (1H, t, J = 6.4 Hz, 1'-H), 7.05-7.45 (15H, m, 6-H and Ar-H).

20 ¹³C NMR (75 MHz, CDCl₃) δ 41.1 (2'-CH₂), 47.3 (5'-CH₂), 71.5 (A₁Ph₂C), 72.9 (3'-CH), 85.5 (1'-CH), 86.8 (4'-CH), 103.0 (5-CH), 126.8 (CITr-CH), 126.9 (CITr-CH), 127.6 (CITr-CH), 128.4 (CITr-CH), 128.5 (CITr-CH), 128.6 (CITr-CH), 129.2 (CITr-CH), 132.3 (CITr-CH), 132.9 (CITr-CH), 134.7 (CITr-C), 140.0 (6-CH), 140.9 (CITr-C), 144.8 (CITr-C), 146.0 (CITr-C), 150.7 (2-C), 163.7 (4-C).

ES⁺ m/z (%) 526 ([M+Na]⁺, 11), 504 ([M+H]⁺, 14), 277 (ClTr⁺, 100). ES m/z (%) 504 (M-H+, 11), 111 (uracil-H+, 100). HRMS (ES⁺) Found [M+H]⁺ 504.1689; C₂₈H₂₇N₃O₄Cl⁺ requires 504.1685. M.p. 129-131°C.

Example 25

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10

5'-Triphenylsilyloxy-2',3'-dideoxydidehydrouridine (25)

To a solution of 2',3'-dideoxydidehydrouridine (0.316 g, 1.50 mmol) in dry pyridine (5 mL) cooled in an ice-salt bath was added drop-wise a solution of triphenylsilyl chloride (0.595 g, 2.02 mmol) in dry pyridine (3 mL). The reaction mixture was kept at 0°C under nitrogen for 2h30. As TLC monitoring evidenced the presence of unreacted starting material, additional triphenylsilyl chloride (0.296 g, 1.00 mmol) in dry pyridine (1 mL) was added. After 1h30 min the reaction was quenched with CH₃OH (50 uL). Removal of the solvent *in vacuo* afforded a crude white gum which was purified by silica gel chromatography (Jones Chromatography Isolute SI column) eluted with $0 \rightarrow 5\%$ CH₃OH in CHCl₃. The title was obtained as a white solid (0.476 g, 68%) from the fractions with R_f = 0.57 (10% CH₃OH/CHCl₃).

¹H NMR (300 MHz, CDCl₃) δ 4.04 (1H, dd, J = 2.2, 11.7 Hz, 5'-H), 4.19 (1H, dd, J = 2.5, 11.7 Hz, 5'-H), 4.78 (1H, dd, J = 1.9, 8.1 Hz, 5-H), 4.98 (1H, m, 4'-H), 5.90 (1H, d, J = 5.7 Hz, 1'-H), 6.33 (1H, dd, J = 1.4, 4.5 Hz, 2'-H), 7.12 (1H, m, 3'-H), 7.40-7.65 (15H, m, Ph-H), 7.80 (1H, d, J = 8.1 Hz, 6-H), 8.99 (1H, bs, 3-NH).

¹³C NMR (75 MHz, CDCl₃) δ 64.7 (5'-CH₂), 87.4 (1'-CH), 90.0 (4'-CH), 102.6 (5-CH),

127.1 (2'-CH), 128.6 (Ph-CH), 131.0 (Ph-CH), 133.3 (Ph-C), 134.9 (3'-CH), 135.8 (Ph-CH), 141.5 (6.0H), 451.3 (2.0H), 463.6 (4.0H)

20 CH), 141.5 (6-CH), 151.2 (2-C), 163.6 (4-C).

ES⁺ m/z (%) 491 ([M+Na]⁺, 36), 119 (100).

HRMS (ES⁺) Found [M+NH₄]⁺ 446.1887; C₂₅H₂₈N₃OSi⁺ requires 446.1894. *M.p.* 73-74°C.

Anal (%) found C 67.75, H 5.04, N 5.84, Cl 1.89;

25 Calcd for C₂₇H₂₇N₂O₄Si, 0.25 HCl C 67.89, H 5.12, N 5.86, Cl 1.86.

Example 26

5'-Pixylamino-2',5'-dideoxyuridine (26)

2'-Amino-2',5'-deoxyuridine (0.231 g, 1.02 mmol) was reacted with pixyl chloride
(0.390g, 1.33 mmol) in dry pyridine (5 mL) at 40°C for 48h. H₂O (10 mL) was added and the crude mixture was extracted with DCM (2 x 15 mL). The organic layers were dried over Na₂SO₄, concentrated in vacuo and purified by flash column chromatography eluting the column (ISOLUTE SI) with a gradient of 0 → 10% CH₃OH in CHCl₃. The fractions with R_f = 0.11 (10% CH₃OH/CHCl₃) yielded the title compound as a white solid (0.118 g, 24%).

¹H NMR (300 MHz, CDCl₃) δ 1.31 (1H, m, 2'-CHH), 2.04 (1H, m, 2'-CHH), 2.29 (1H, dd, J = 4.3, 13.8 Hz, 5'-CHH), 2.58 (1H, m, 5'-CHH), 3.84 (2H, m, 3'-H and 4'-H), 5.69 (1H, d, J = 8.1 Hz, 5-H), 6.21 (1H, t, J = 6.5 Hz, 1'-H), 7.03-7.53 (14H, m, 6-H and Ar-H). ¹³C NMR (75 MHz, CDCl₃) δ 39.4 (2'-CH₂), 42.3 (5'-CH₂), 72.1 (3'-CH), 76.9 (pixyl-C),

- 84.7 (1'-CH), 86.1 (4'-CH), 103.1 (5-CH), 116.8 (pixyl-CH), 116.9 (pixyl-CH), 123.3 (pixyl-C), 123.6 (pixyl-C), 123.97 (pixyl-CH), 124.03 (pixyl-CH), 127.4 (pixyl-CH), 127.6 (-CH), 128.2 (pixyl-CH), 130.20 (pixyl-CH), 130.24 (pixyl-CH), 131.1 (pixyl-CH), 131.5 (pixyl-CH), 140.3 (6-CH), 148.0 (pixyl-C), 150.5 (2-C), 151.4 (pixyl-C), 151.9 (pixyl-C), 163.5 (4-C).
- ES⁺ m/z (%) 506 ([M+Na]⁺, 4), 257 (Pixyl⁺ that is C₁₉H₁₃O⁺, 100). ES⁻ m/z (%) 482 (M-H⁺, 100). HRMS (ES⁺) Found [M+H]⁺ 484.1871; C₂₈H₂₆N₃O₅⁺ requires 484.1867. M.p. 117-119°C.

Anal calcd for C₂₈H₂₅N₃O₅ (%): 1.58 HCl C 62.15, H 4.95, N 7.7;7 found C: 62.07, H 4.66, N 7.50.

Example 27

5'-O-trityl-2'-deoxyuridine (27)

2'-deoxyuridine (4.00 g, 17.5 mmol) and triphenylmethyl chloride (5.37 g, 19.25 mmol) were stirred in anhydrous pyridine (70 ml) at 50 °C overnight. Additional triphenylmethyl chloride (1.00 g, 3.59 mmol) was added after 18 hours, and the mixture was stirred for a further 4 hours at 50 °C. The reaction mixture was then poured into ice-H₂O (300 ml) and stirred vigorously. The precipitate was extracted with EtOAc (3×100 ml). The organic solution was then washed with 0.5 M HCl (4×100 ml), dried with MgSO₄ and filtered. The filtrate was then washed further with EtOAc, which was then evaporated, and finally with DCM. Solvent was removed using a Buchi rotary evaporator, and finally with the vacuum pump.

White solid (7.69 g, 93%).

15 <u>Example 28</u>

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3'-O-Mesyl-5'-O-trityl-2'-deoxyuridine (28)

Methanesulphonyl chloride (0.173 ml, 2.24 mmol) was added to a solution of (8) (0.30 g, 0.64 mmol) in anhydrous pyridine (5 ml) with ice-bath cooling. The mixture was stirred for 4 hours at room temperature. After this time, ice-water (1 ml) was added; the mixture was stirred for 5 minutes, then poured into ice-water (30 ml) and filtered. The precipitate was dissolved in CHCl₃ (30 ml), the solution was washed with 0.5 M HCl (10 ml) and

water (3×10 ml), dried (MgSO₄), filtered, and reduced in vacuo which gave the title product as a yellow/orange solid (0.29 g, 83%).

Example 29

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2, 3'-Anhydro-5'-O-trityl-2', 3'-dideoxyuridine (29)

DBU (1.00 ml, 7.12 mmol) and compound 28 (3.55 g, 6.48 mmol) were stirred in DCM (25 ml) over 30 hours. The mixture was washed with water (2×30 ml), the organic layer was dried (MgSO₄), filtered and reduced in vacuo. The residue was purified by column chromatography (5% MeOH/CHCl₃) which gave the title product as a white solid (2.42 g, 82%).

Example 30

15 3'-Azido-5'-O-trityl-2', 3'-dideoxyuridine (30)

Lithium fluoride (0.145 g, 5.61 mmol) was suspended in DMF (3 ml) and heated to 105 °C with stirring. To the stirred suspension was added N,N,N',N'tetramethylethylenediamine (5 ml) followed by azidotrimethylsilane (0.64 g, 5.61 mmol).

After stirring for an hour, compound 29 (1.41 g, 3.11 mmol) dissolved in N,N-

dimethylformamide (2 ml) was added, and the reaction was allowed to proceed for 20 hours at 110 °C. The mixture was cooled, poured into CHCl₃ (110 ml) and filtered through Celite. The solvent was removed under reduced pressure and the residue (brown oil) was taken in EtOAc (100 ml). The organic phase was washed with water (4×180 ml), dried (MgSO₄), filtered and concentrated. The concentrated mixture was

purified by column chromatography (3% MeOH/CHCl₃) which gave the title product as an orange solid (0.996 g, 65 %).

Example 31

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3'-Amino-5'-O-trityl-2',3'-dideoxyuridine (31)

Lindlar's catalyst (20 mg) was added to compound 30 (0.10 g, 0.20 mmol), and was then suspended in ethanol (5 ml). Air was removed from the flask by flushing with nitrogen several times. The nitrogen was then removed and replaced with hydrogen.

The mixture was stirred for 5 hours, and then filtered through Celite. Fresh Lindlar's catalyst (20 mg) was added to the filtrate. The flask was flushed with nitrogen and then hydrogen as previously, and the reaction was left stirring for another 3 hours. The reaction mixture was filtered through Celite. The solvent was evaporated and the concentrated solution was purified by column chromatography (MeOH/DCM 2% → 10 %) which gave the title product as a white solid (0.065 g, 70%). R_f: 0.3 in DCM/MeOH 90:10.

¹H-NMR (300 MHz, CDCl₃): δ 7.97 (d, J=8.2 Hz, 1H, H-6), 7.47–7.27 (m, 15H, H-aromatic), 6.20 (q, J=3.3 Hz, 1H, H-1') 5.41 (d, J=8.1 Hz, 1H, H-5), 3.73–3.38 (m, 5H, H-3', H-4', H-5'), 2.44 – 2.18 (m, 2H, H-2').

¹³C-NMR (75 MHz, CDCl₃): δ 163.5 (C-4), 150.5 (C-2), 143.7 (C-7'), 140.7 (C-6), 129.1 (C-8'), 128.5 (C-9'), 127.8 (C-10'), 102.2 (C-5), 87.9 (C-6'), 87.2 (C-4'), 85.2 (C-1'), 62.2 (C-5'), 50.7 (C-3'), 42.7 (C-2').

LRMS: (ES+ mode):m/z=491.7 [(M+Na)⁺, 45%]; m/z=243.2 [(Tr)⁺, 100%].

HRMS: (ES+ mode): found 492.1902; required 492.1899 for $C_{28}H_{27}N_3O_4Na$ [M Na][†]. Microanalysis calculated for $C_{28}H_{27}N_3O_4 \times 0.5$ H₂O:

C 70.28, H 5.90, N 8.78%; found: C 70.64, H 5.92, N 8.41%.

Example 32

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3'-Acetylamino-5'-O-trityl-2', 3'-dideoxyuridine (32)

Compound 31 (0.10 g, 0.213 mmol) was suspended in DCM (5 ml), and to this was added acetic anhydride (0.047 g, 0.044 ml, 0.469 mmol) and triethylamine (0.065 ml, 0.469 mmol). The mixture was stirred at room temperature for 3 hours. After this time the solvent was evaporated to give a white solid. The product was purified by column chromatography (MeOH/DCM $2\% \rightarrow 6\%$), and evaporation of the solvent gave the title compound as a white solid (0.103 g, 95%).

10 R_f: 0.45 in DCM/MeOH 90:10.

¹H-NMR (300 MHz, CDCl₃): δ 9.82 (s, 1H, N-H), 7.83 (d, J=8.2 Hz, 1H, H-6), 7.46–7.28 (m, 16H, H-aromatic) 6.94 (s, 1H, N-H), 6.34 (t, J=6.3 Hz, 1H, H-1'), 5.39 (d, J=8.1 Hz, 1H, H-5), 4.79–4.72 (m, 1H, H-3'), 4.07 (s, 1H, H-4') 3.59–3.47 (m, 2H, H-5'), 2.50–2.32 (m, 2H, H-2'), 2.04 (s, 3H, H-12')

¹³C-NMR (75 MHz, CDCl₃): δ 163.6 (C-4), 154.9 (C-2), 143.6 (C-7'), 140.5 (C-6), 129.1 (C-8'), 128.5 (C-9'), 127.9 (C-10'), 103.1 (C-5), 88.1 (C-6'), 87.3 (C-1'), 85.2 (CH, C-4'), 62.1 (C-5'), 50.9 (C-3'), 38.8 (C-2')

LRMS: (ES+ mode): m/z = 533.8 [(M+Na)⁺, 20%].

HRMS: (ES+ mode): Found 534.2009; required 534.2005 for $C_{30}H_{29}N_3O_5Na$ [M+Na]⁺ Microanalysis calculated for $C_{30}H_{29}N_3O_5 \times 1.0$ HCl $\times 1.0$ H₂O C 63.66, H 5.70, N 7.42%; found C 63.20, H 5.15, N 7.11%.

Example 33

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Diphenyl(pyridin-2-yl)methanol (33)

A solution of 2-bromopyridine (5 g, 0.032 mol) in dry THF (15 0mL) was cooled to -70 °C. To this cooled solution was added n-BuLi (2.8 M, 12.4 mL, 0.034 mol) over a period of 20 min and allowed to stir for 2h under N₂ atmosphere. A solution of benzophenone (5.8 g, 0.032 mol) in dry THF (50 mL) was added to the solution at the same temperature over a period of 30 min. The reaction mixture was warmed slowly to RT and allowed to stir another 5h at RT. The reaction mixture was concentrated under vacuum and the residue was washed with petrolieum ether. The organic layer was filtered and the filtrate was concentrated under vacuum to give the title compound (8 g, 95%).

Example 34

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2-[Chloro (diphenyl)methyl]pyridine hydrochloride (34)

To a mixture of diphenyl(pyridin-2-yl)methanol (4 g, 0.015 mol) in thionylchloride (50 mL) was added acetylchloride (15 mL, 0.195 mol) at RT and heated to 50 °C for 48h. The reaction mixture was concentrated under vacuum and the residue was azeotroped with dry benzene (100 mL x 2) to give the title compound as the hydrochloride salt (4.4 g, >95%).

Example 35

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4-[Hydroxy(diphenyl)methyl]benzonitrile (35)

The procedure described in example 49 was followed but using 4-bromobenzonitrile (5g, 0.027mol) instead of 2-bromopyridine which gave the title compound (7.5g, 94%).

Example 36

4-[Chloro(diphenyl)methyl]benzonitrile (36)

To a mixture of 4-[hydroxy(diphenyl)methyl] in dry toluene (60 mL) was added acetylchloride (3 mL) at RT and heated to 50 °C for 12h. The reaction mixture was concentrated under vacuum. The residue was recrystallized from pet. ether to give the product (1.7 g, 40%).

Example 37

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10 Diphenyl (pyrimidin-5-yl) methanol (37)

A solution of 5-bromopyrimidine (10 g, 0.063 mol) in a mixture of dry THF (150 mL) and hexane (50 mL) was cooled to -100 °C. To this cooled solution was added n-BuLi (4 g, 21 mL, 0.062 mol) over a period of 30 min and stirred for another 30 min. A solution of benzophenone (11.5 g, 0.063 mol) in dry THF (50 mL) was added to this at the same temperature over a period of 30min. The reaction mixture was warmed slowly to RT and allowed to stir another 1h at RT. The reaction was quenched with cold water (200 mL), ethyl acetate was added and the organic layer was separated. The organic layer was dried, concentrated and the crude product was purified by column chromatography on silica gel (up-to 25% ethyl acetate in pet. ether) to give the product (8g). TLC: Pet. ether/EtOAc, 1:1, R_f=0.3

Example 38

2,2,2-Triphenylethanol (38)

To a suspension of LAH (3.9 g, 0.104 mol) in dry THF (200 mL) was stirred at 0 °C for 20 min. A solution of 2,2,2-triphenylacetic acid (10 g, 0.034 mol) in dry THF (50 mL) was added in a drop-wise manner. The reaction mixture was stirred at RT overnight. Excess LAH was quenched with 1.5 N HCl and the reaction mixture was further stirred for 2h at RT. The reaction mixture was filtered through celite, washed with ethyl acetate and the filtrate was concentrated under vacuum. The crude product was purified by column chromatography on silica gel (4% ethyl acetate in pet. ether) to give the title compound (4.6 g, 48%). TLC: Pet. ether/EtOAc, 7:3, R_f=0.2

Example 39

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15 3,3,3-Triphenylpropan-1-ol (39)

To a magnetically stirred suspension of LAH (8.3 g, 0.219 mol) in dry THF (50 mL) was added a solution of 3,3,3-triphenylpropionic acid (9.5 g, 0.0314 mol) over a period of 30 min at 0 °C. The reaction mixture was allowed to stir at RT for 14h. The reaction mixture was cooled and excess LAH was quenched with 20% NaOH solution (50 mL). The reaction mixture was passed through celite, washed with THF and the filtrate was concentrated under vacuum. The residue was washed with pet. ether and dried which gave the title compound (8g, >85%).

TLC: Pet. ether/EtOAc, 7:3, R_f=0.2

Example 40

1,1,2-Triphenylethanol (40)

To a suspension of Mg (1.7 g, 0.07 mol) in dry ether (25 mL) was added a solution of benzyl bromide (10 mL, 1.5 equ.) in dry ether (25 mL) drop-wise and allowed to stir at RT for 1h. By the time all magnesium was dissolved and the reaction mixture was cooled to 0 °C. To this was added a solution of benzophenone (10 g, 0.05 mol) in dry ether (25 mL) and allowed to stir at RT for 5h. The progress of the reaction was followed by TLC and when it was ready the reaction mixture was quenched with saturated NH₄Cl solution, extracted with ether (100 mL), washed with brine, dried and concentrated under vacuum. The crudeproduct was purified by column chromatography on silica gel (10% ethyl acetate in pet. ether) to give the title compound (9.6 g, 65%) as a white solid.

TLC: Pet. ether/EtOAc, 9:1, R_f=0.4

Example 41

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4-Trityloxy-but-2-en-1-ol (41)

Trityl chloride (557 mg; 2 mmol) Et₃N (0.306 ml; 2.2 mmol) and DMAP (10 mg; 0.08 mmol) were added to a emulsion of cis-2-buten-1,4-diol (1.76 g; 20 mmol) in DCM (10 ml). The mixture was stirred at room temperature under atmosphere of nitrogen for 24 hours. After such period of time the complete disappearance of trityl chloride was observed by TLC (EtOAc/Hexane 50:50). DCM (20 ml) and water (10 ml) were added to the mixture. The phases were separated and the organic layer was washed with water (10 ml) and brine (10 ml). The solvent was dried over MgSO₄ and evaporated under reduced pressure affording a residue (white oil) which was purified by flash

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chromatography using Hexane/EtOAc 70:30 → 40:60 as gradient which gave the title product as a colourless oil (563 mg, 81%).

Example 42

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Trans-2-buten-1,4-diol (42)

2-Butyn-1,4-diol (1g; 11.64 mmol) was dissolved in dry THF (25 ml) under atmosphere of nitrogen. The solution was cooled to -78 °C with a dry-ice/acetone bath. A cold solution of LAH in THF 1M (12.7 ml; 12.7 mmol) was added with a syringe. The reaction was left worm to room temperature in 4 hours. The disappearance of the starting alkyne was observed by TLC (Hexane/EtOAc 30:70); then the solution was cooled to -0 °C with an ice bath and the quenched with NaOH 1M, until no gas was developed. The pH was adjusted to 8 with HCl 1M and then silica was added to the solution. The solvents were removed under reduced pressure and the residue was loaded into a chromatographic column and purified using Hexane/EtOAc 30:70 as eluent which gave the title compound as a colourless oil (817 mg, 79%). R_f: 0.11 in Hexane/EtOAc 30:70 (PMA)

¹H-NMR (300 MHz, CD₃OD): δ 5.83 (bs; 2H; H-2+H-3); 4.07 (d; J=3.57 Hz; 4H; H-1+H-4)

¹³C-NMR (75 MHz, CD₃OD): δ 131.7 (C-2 & C-3); 63.4 (C-1 & C-4) LRMS (ES+): m/z 111.0 [M+Na]⁺ 100%.

Example 43

25 (E)-4-(Trityloxy)but-2-en-1-ol (43)

A solution of Trityl chloride (500 mg; 1.81 mmol), TEA (0.277 ml; 1.99 mmol) and DMAP (8.8 mg; 0.072 mmol) in dry DCM (5 ml) was added with a syringe to a solution of the diol (10) (800 mg; 9.07 mmol) in DCM (15 ml). The mixture was stirred at room temperature for 1 hour and 30 minutes, then other TrCl, TEA and DMAP (half quantities

than before) were added. The reaction was stirred at the same temperature until TLC (Hexane/EtOAc 50:50) showed complete disappearance of Trityl chloride. After 1.5 hours water (20 ml) was added and the reaction was stirred for few minutes, then the phases were separated. The organic layer was washed with water (25 ml) and brine (25 ml). The solvent was dired over MgSO₄ and evaporated to afford a crude oil which was purified by flash chromatography using Hexane/EtOAc 50:50 as eluent which gave the title compound as a colourless oil, 637 mg, 71%. R_f: 0.72 in Hexane/EtOAc 50:50 (UV/PMA).

¹H-NMR (300 MHz, CDCl₃): δ 7.54-7.51 (m; 6H; H-7); 7.39-7.27 (m; 9H; H-8+H-9); 6.12-6.03 (m; 1H; H-3); 5.91-5.83 (m; 1H; H-2); 4.24 (bs; 2H; H-4); 3.71-3.71 (m; 2H; H-1).

¹³C-NMR (75 MHz, CDCl₃): δ 144.6 (C-6); 130.5 (C-3 & C-2); 129.0 (C-8); 128.7 (C-7); 127.4 (C-9); 87.3 (C-5); 64.5 (C-4); 63.8 (C-1).

LRMS (ES+): m/z 331.2 [M+H]⁺ 100%.

Example 44

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5- Trityloxypentanol (44)

The procedure described in example 41 was followed but using 1,5-pentandiol (376 mg, 3.6 mmol) as alcohol instead of cis-2-buten-1,4-diol in the reaction with trityl chloride, which gave the title compound (300 mg, 24%).

Example 45

25 5-(Tritylamino)-pentan-1-ol (45)

The title compound (24%) was prepared as described in example 41 but using 5-aminopentanol instead of cis-2-buten-1,4-diol in the reaction with trityl choride.

5 Biological Examples

Example B1

Malaria whole cell assays

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Parasite cultures

Two strains of *P.falciparum* are used in this study: The drug sensitive NF54 (an airport strain of unknown origin) and K1 (Thailand, chloroquine and pyrimethamine resistant). The strains are maintained in RPMI-1640 medium with 0.36 mM hypoxanthine supplemented with 25 mM HEPES, 25 mM NaHCO₃, neomycin (100 U/ml) and Albumax^R (lipid-rich bovine serum albumin) (GIBCO, Grand Island, NY) (5g/l), together with 5% washed human A+ erythrocytes. All cultures and assays are conducted at 37°C under an atmosphere of 4% CO₂, 3% O₂ and 93% N₂. Cultures are kept in incubation chambers filled with the gas mixture. Subcultures are diluted to a parasitaemia of 0.1-0.5% and the medium changed daily.

Drug sensitivity assays

Antimalarial activity is assessed using an adaptation of the procedures described by Desjardins et al.(Antimicrob. Agents Chemother. 16(6):710-8, 1979), and Matile and Pink (In: Lefkovits, I. and Pernis, B. (Eds.). Immunological Methods. Academic Press, San Diego, pp. 221-234, 1990).

Stock drug solutions are prepared in 100% DMSO (dimethylsulfoxide) at 10 mg/ml, unless otherwise suggested by the supplier, and heated or sonicated if necessary. After use the stocks are kept at –20°C. The compound is further diluted to the appropriate concentration using complete medium without hypoxanthine.

Assays are performed in sterile 96-well microtiter plates, each well containing 200 μl of parasite culture (0.15% parasitemia, 2.5% hematocrit) with or without serial drug solutions. Seven 2-fold dilutions are used covering a range from 5 μg/ml to 0.078 μg/ml.

For active compounds the highest concentration is lowered (e.g. to 100 ng/ml), for plant extracts the highest concentration is increased to 50 μ g/ml. Each drug is tested in duplicate and repeated once for active compounds showing an IC₅₀ below 0.5 μ g/ml. After 48 hours of incubation at 37°C, 0.5 μ Ci ³H-hypoxanthine is added to each well.

Cultures are incubated for a further 24 h before they are harvested onto glass-fiber filters and washed with distilled water. The radioactivity is counted using a BetaplateTM liquid scintillation counter (Wallac, Zurich, Switzerland). The results are recorded as counts per minute (CPM) per well at each drug concentration and expressed as percentage of the untreated controls. From the sigmoidal inhibition curves IC₅₀ values are calculated.

Primary screen

K1 strain is used. The compounds are tested at 7 concentrations (5000 to 78 ng/ml). Artemisinin and chloroquine are included as reference drugs.

If the IC₅₀ is >5 μg/ml, the compound is classified as inactive
If the IC₅₀ is 0.5-5 μg/ml, the compound is classified as moderately active
If the IC₅₀ is <0.5 μg/ml, the compound is classified as active and is further evaluated using two strains, K1 and NF54. A new range of concentrations is chosen depending on the IC₅₀ determined (e.g. 100 to 1.56 ng/ml) and the assay is carried out 2x
independently.

The standard drugs are chloroquine and artemisinin which are run in the same assay. The IC₅₀ values for chloroquine are 2.9 ng/ml for NF54 and 48 ng/ml for K1; for artemisinin 1.9 ng/ml for NF54 and 0.8 ng/ml for K1.

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Secondary screen

Test compounds are tested against a panel of say, 14 different of different origin and some show resistances to chloroquine and/or pyrimethamine. If the range of the IC₅₀ values for the 14 strains is within a factor 3-5 x then the tested compound is considered not to show cross resistance.

Example B2

Malaria enzyme assays

Inhibition of Plasmodium falciparum dUTPase

Chemicals

2′-dUTP, was purchased from Pharmacia. MgCl₂, BSA, and the pH indicator cresol red were from Sigma. The buffer N, N-bis (2-hydroxyethyl) glycine (BICINE) was obtained from USB (United States Biochemical), Ohio. All the concentrations of nucleotides were calculated spectrophotometrically (HP-8453, Hewlett Packard) at 280 nm, using the extinction coefficient (ε_{280 nm}=1.75 ml mg⁻¹cm⁻¹). Other chemicals used in these experiments were of the highest quality available.

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Cloning of the PFdut gene

Conserved motifs of the human dUTPase enzyme were used as query to identify the *PFdut* gene in the www. tigr.org database of the *Plasmodium falciparum* 3D7 strain. The entire coding sequence was amplified by the PCR using as template cDNA and as primers the oligonucleotides *ATG-PFdut* (CATATGCATTTAAAAATTGTATGTCTG) and *TGA-PFdut* (GGATCCTCAATATTTATTATCGATGTCGATC) which were designed so that *Ndel* and *BamHl* restriction sites were introduced at the 5' and 3' ends for convenient cloning in the expression vector pET11 (*Stratagene*). The amplified product was cloned into pGEMT (*Promega*) and propagated in *E. coli* XL1B cells. In order to confirm the correct sequence after amplification, sequencing was performed using an Applied Biosystems Automated Sequencer, at the Analytical Services of the Instituto de Parasitología y Biomedicina "López Neyra". These Services also supplied the oligonucleotides designed for the sequencing

25 P. falciparum dUTPase overexpression and purification

Recombinant *P. falciparum* dUTPase was purified from *E. coli* BL21 (DE3) cells transformed with pET-*PFdut*. Pellets from a liter of culture were resuspended in a solution consisting of buffer A (20 mM MES pH 5.5, 50 mM NaCl, 1 mM DTT) plus the protease inhibitors 1 mM PMSF, 20 μg/ml leupeptin and 1 mM benzamidine. Purification was carried out in a cold room (4°C). The soluble crude extract was obtained by sonication in a Vibra-cell (Sonics and Materials Inc. Danbury, Connecticut, USA) and centrifugation at 14000xg. The extract was loaded onto a phosphocellulose column (Whatman) pre-equilibrated with buffer A at a flow rate of 1 ml/min. After washing the

column with 100 ml of buffer A, elution was performed using a linear NaCl gradient of 50 to 1000 mM. Peak fractions with a low concentration of contaminating protein, as judged by 15% SDS-PAGE gels, were pooled and then loaded and chromatographed on a *Superdex 200* column at a flow rate of 0.5 ml/min. The column was equilibrated with buffer B (50 mM Bicine, 1 mM DTT, 10 mM MgCl₂). Peak fractions were pooled and concentrated to about 5 mg/ml by ultrafiltration in a Centripep tube (*Amicon*) and stored at –80°C.

Kinetic measurements

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Nucleotide hydrolysis was monitored by mixing enzyme and substrate with a rapid kinetic accessory (Hi-Tech Scientific) attached to a spectrophotometer (Cary 50) and connected to a computer for data acquisition and storage. Protons, released through the hydrolysis of nucleotides, were neutralised by a pH indicator in a weak buffered medium with similar pK_a and monitored spectrophotometrically at the absorbance peak of the basic form of the indicator. Absorbance changes were kept within 0.1 units. The indicator/buffer pair used was cresol red/BICINE (2mM/50μM, pH 8, 573 nm). The measurements were performed at 25°C, and the solutions were previously degassed. Assays contained 30 nM purified recombinant enzyme, 50 μM dUTP, 5mM MgCl₂ and 2.5mM DTT, 1.25mg/ml BSA and 100mM KCl. Indicator absorbance changes corresponding to complete hydrolysis of nucleotides were recorded in the computer, and the kinetic parameters *V*_{mex} and *K*_{mapp} (or slope) were calculated by fitting the data to the integrated Michaelis-Menten equation (Segel, 1975). [dUMP]/t=V_{max}-K_{map}/t In [dUTP]/([dUTP]-[dUMP])

Solutions of potential inhibitors were prepared at 10mg/ml and tested routinely at concentrations of 2, 10, and $50\mu g/\mu l$. A wider range of concentrations was further tested when necessary for K_i determination. The different apparent K_m values attained were plotted against inhibitor concentration and K_i values were obtained according to the following equation:

$$K_{map} = \underline{K}_{m} [1] + K_{m}$$

$$K_{i}$$

Example B3

Human dUTPase assay

Human recombinant dUTPase was purified from *E. coli* BL21 (DE3) cells

transformed with pETHudut (Dr. P.O. Nyman, Lund University). Purification was accomplished as described for the dUTPase above except that the last step in
Superdex 200 was omitted. Likewise, conditions for enzyme assays were the same as described above except that the enzyme concentration was 50nM.

10 Example B4

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Trypanosoma brucei whole cell assays

Parasite cultures

Three strains of *T. brucei* spp. are used in this study: (a) *Trypanosoma brucei rhodesiense* STIB 900, a clone of a population isolated in 1982 from a patient in Tanzania which is known to be susceptible to all currently used drugs; (b) *Trypanosoma brucei gambiense* STIB 930, a derivative of strain TH1/78E (031) isolated in 1978 from a patient in Ivory Coast which is known to be sensitive to all drugs used, and (c) *Trypanosoma brucei brucei* STIB 950, a clone of a population isolated in 1985 from a bovine in Somalia which shows drug resistance to diminazene, isometamidium and quinapyramine.

The bloodstream form trypomastigotes of the strains a and c are maintained in MEM medium with Earle's salts supplemented with 25 mM HEPES, 1g/l additional glucose, 1% MEM non-essential amino acids (100x), 0.2 mM 2-mercaptoethanol, 2mM Napyruvate, 0.1mM hypoxanthine and 15% heat inactivated horse serum. The bloodstream form trypomastigotes of strain b are maintained in MEM medium with Earle's salts supplemented with 25 mM HEPES, 1g/l additional glucose, 1% MEM non-essential aminoacids (100x), 0.2 mM 2-mercaptoethanol, 2mM Na-pyruvate, 0.1mM hypoxanthine, 0.05mM bathocuproine disulphonic acid, 0.15mM L-cysteine and 15% heat inactivated pooled human serum.

All cultures and assays are conducted at 37°C under an atmosphere of 5% CO₂ in air.

Drug sensitivity assays

Stock drug solutions are prepared in 100% DMSO (unless otherwise suggested by the supplier) at 10 mg/ml, and heated or sonicated if necessary. After use the stocks are kept at -20°C. For the assays, the compound is further diluted to the appropriate concentration using complete medium.

Assays are performed in 96-well microtiter plates, each well containing 100 μl of culture medium with 8 x 10 ³ bloodstream forms with or without a serial drug dilution. The highest concentration for the test compounds is 90 μg/ml. Seven 3-fold dilutions are used covering a range from 90 μg/ml to 0.123 μg/ml. Each drug is tested in duplicate and each assay is repeated at least once. After 72 hrs of incubation the plates are inspected under an inverted microscope to assure growth of the controls and sterile conditions.

15 10μl of Alamar Blue (12.5 mg resazurin dissolved in 100 ml distilled water) are now added to each well and the plates incubated for another 2 hours. Then the plates are read with a Spectramax Gemini XS microplate fluorometer (Molecular Devices Cooperation, Sunnyvale, CA, USA) using an excitation wave length of 536 nm and an emission wave length of 588 nm. Data are analysed using the microplate reader software Softmax Pro (Molecular Devices Cooperation, Sunnyvale, CA, USA).

Primary screen

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The preliminary screen uses the *Trypanosoma b. rhodesiense* strain. The compounds are tested at 7 concentrations (drug concentrations ranging from 90 μ g/ml to 0.123 μ g/ml in 3-fold dilutions).

If the IC $_{50}$ is >3 μ g/ml, the compound is classified as inactive If the IC $_{50}$ is 0.2-3 μ g/ml, the compound is classified as moderately active If the IC $_{50}$ is <0.2 μ g/ml, the compound is classified as active

The standard drug is melarsoprol which is run in the same assay; the IC₅₀ for melarsoprol is 1.6 ng/ml.

Secondary screen

Active compounds (IC_{50} <0.2 μ g/ml) are tested against the *Trypanosoma brucei* gambiense STIB 930 and the drug resistant *T. b. brucei* STIB 950 following the same protocol as described above.

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The standard drug is melarsoprol which is run in the same assay; the IC $_{50}$ for melarsoprol is 4.2 ng/ml for STIB 930 and 2.8 ng/ml for STIB 950

Example B5

10 Trypanosoma cruzi whole cell assays

Trypanosoma cruzi cell cultures:

The *Trypanosoma cruzi* Tulahuen C2C4 strain, containing the -galactosidase (Lac Z) gene, is used. The plasmid construct by Dr. S. Reed was obtained from Dr. F. Buckner, Seattle, as epimastigotes in LIT medium.

The infective amastigote and trypomastigote stages are cultivated in L-6 cells (rat skeletal myoblast cell line) in RPMI 1640 supplemented with 2 mM L-glutamine and 10% heat-inactivated foetal bovine serum in 12.5 cm² tissue culture flasks. Amastigotes develop intracellularly, differentiate into trypomastigotes and leave the host cell. These trypomastigotes infect new L-6 cells and are the stages used to initiate an infection in the assay. All cultures and assays are conducted at 37°C under an atmosphere of 5% CO₂ in air.

25 Drug sensitivity assays

Stock drug solutions are prepared in 100% DMSO (dimethylsulfoxide) unless otherwise suggested by the supplier at 10 mg/ml, and heated or sonicated if necessary. The stocks are kept at -20°C. For the assays, the compound is further diluted to the appropriate concentration using complete medium.

Assays are performed in sterile 96-well microtiter plates, each well containing 100 μ l medium with 2x10³ L-6 cells. After 24 hours 50 μ l of a trypanosome suspension containing 5 x 10³ trypomastigote bloodstream forms from culture are added to the

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wells. 48 hours later the medium is removed from the wells and replaced by 100 μ l fresh medium with or without a serial drug dilution. At this point the L-6 cells should be infected with amastigotes and no free trypomastigotes should be in the medium. Seven 3-fold dilutions are used covering a range from 90 μ g/ml to 0.123 μ g/ml. Each drug is tested in duplicate. After 96hours of incubation the plates are inspected under an inverted microscope to assure growth of the controls and sterility.

Then the substrate CPRG/ Nonidet (50 I) is added to all wells. A colour reaction will become visible within 2-6 hours and can be read photometrically at 540nm. Data are transfered into a graphic programme (e.g. EXCEL), sigmoidal inhibition curves determined and IC₅₀ values calculated.

Primary screen

Benznidazole is used as the reference drug and shows an IC50 value of 0.34 µg/ml.

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If the IC $_{50}$ is > 30 µg/ml, the compound is classified as inactive. If the IC $_{50}$ is between 2 and 30µg/ml, the compound is classified as moderately active. If the IC $_{50}$ is < 2 µg/ml, the compound is classified as active.

20 Example B6

Leishmaniasis: macrophage in vitro screening model

Parasite and cell cultures

The *Leishmania.donovani* strain MHOM/ET/67/L82 obtained from Dr. S. Croft, London) is used. The strain is maintained in the Syrian Golden hamster. Amastigotes are collected from the spleen of an infected hamster Amastigotes are grown in axenic culture at 37°C in SM medium (Cunningham I., J. Protozool. 24, 325-329, 1977) at pH 5.4 supplemented with 10% heat-inactivated foetal bovine serum under an atmosphere of 5% CO₂ in air.

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Primary peritoneal macrophages from NMRI mice are collected 1 day after a macrophage production stimulation with an i.p injection of 2ml of a 2% potato starch

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suspension (FLUKA, Switzerland) All cultures and assays are done at 37°C under an atmosphere of 5% CO₂ in air.

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Drug sensitivity assays

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Stock drug solutions are prepared in 100% DMSO (unless otherwise suggested by the supplier) at 10 mg/ml, and heated or sonicated if necessary. After use the stocks are kept at -20°C. For the assays, the compound is further diluted in serum-free culture medium and finally to the appropriate concentration in complete medium.

Assays are performed in sterile 16-well chamber slides (LabTek, Nalgene/Nunc Int.) To each well 100 µl of a murine macrophage suspension (4 x 10⁵/ml) in RPMI 1640 (containing bicarbonate and HEPES) supplemented with 10% heat inactivated fetal bovine serum is added. After 24 hrs 100 µl of a suspension containing amastigotes (1.2 x 10⁶/ml) is added resulting in a 3:1 ratio of amastigotes/macrophages. The amastigotes are harvested from an axenic amastigote culture and suspended in RPMI/FBS. 24 hrs later, the medium containing free amastigotes is removed, washed 1x and replaced by fresh medium containing four 3-fold drug dilutions. In this way 4 compounds can be tested on one 16-well tissue culture slide. Untreated wells serve as controls. Parasite growth in the presence of the drug is compared to control wells. After 4 days of incubation the culture medium is removed and the slides fixed with methanol for 10 min followed by staining with a 10% Giemsa solution. Infected and non-infected macrophages are counted for the control cultures and the ones exposed to the serial drug dilutions. The infection rates are determined. The results are expressed as % reduction in parasite burden compared to control wells, and the IC50 calculated by linear regression analysis.

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Primary screen

The compounds are tested in duplicate at 4 concentrations ranging from 9 to 0.3 μ g/m. If the IC50 is below 0.3 μ g/ml then the range is changed to 1 to 0.03 μ g/ml. Miltefosine is used as the reference drug and shows an IC₅₀ value of 0.325 μ g/ml (0.22 – 0.42

30 μ g/ml; n=4)

If the IC $_{50}$ is higher than 10 μ g/ml, the compound is classified as inactive. If the IC $_{50}$ is between 2 and 10 μ g/ml, the compound is classified as moderately active.

If the IC $_{50}$ is < 2 μ g/ml, the compound is classified as active and is further evaluated in a secondary screening.

Drug sensitivity assays

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Stock drug solutions are prepared in 100% DMSO (dimethylsulfoxide) unless otherwise suggested by the supplier at 10 mg/ml, and heated or sonicated if necessary. The stocks are kept at –20°C. For the assays, the compound is further diluted to the appropriate concentration using complete medium.

Assays are performed in sterile 96-well microtiter plates, each well containing $100~\mu l$ medium with $2x10^3$ L-6 cells. After 24 hours $50~\mu l$ of a trypanosome suspension containing $5~x~10^3$ trypomastigote bloodstream forms from culture are added to the wells. 48 hours later the medium is removed from the wells and replaced by $100~\mu l$ fresh medium with or without a serial drug dilution. At this point the L-6 cells should be infected with amastigotes and no free trypomastigotes should be in the medium. Seven 3-fold dilutions are used covering a range from $90~\mu g/m l$ to $0.123~\mu g/m l$. Each drug is tested in duplicate. After 96hours of incubation the plates are inspected under an inverted microscope to assure growth of the controls and sterility.

Then the substrate CPRG/ Nonidet (50 I) is added to all wells. A colour reaction will become visible within 2-6 hours and can be read photometrically at 540nm. Data are transferred into a graphic programme (e.g. EXCEL), sigmoidal inhibition curves determined and IC₅₀ values calculated.

Primary screen

25 Benznidazole is used as the reference drug and shows an IC₅₀ value of 0.34 μg/ml.

If the IC $_{50}$ is > 30 µg/ml, the compound is classified as inactive. If the IC $_{50}$ is between 2 and 30µg/ml, the compound is classified as moderately active. If the IC $_{50}$ is < 2 µg/ml, the compound is classified as active.

Example B7

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Leishmania donovani, axenic amastigote assay

Parasite and cell cultures:

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The *Leishmania donovani* strain MHOM/ET/67/L82) is used. The strain is maintained in the hamster. Amastigotes are collected from the spleen of an infected hamster and adapted to axenic culture conditions at 37°C. The medium is a 1:1 mixture of SM medium (Cunningham I., J. Protozool. 24, 325-329, 1977) and SDM-79 medium (Brun, R. & Schönenberger, M., Acta Trop. 36, 289-292, 1979) at pH 5.4 supplemented with 10% heat-inactivated FBS under an atmosphere of 5% C0₂ in air.

10 Drug sensitivity assays

Stock drug solutions are prepared in 100% DMSO (unless otherwise suggested by the supplier) at 10 mg/ml, and heated or sonicated if necessary. After use the stocks are kept at -20°C. For the assays, the compound is further diluted to the appropriate concentration using complete medium.

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Assays are performed in 96-well microtiter plates, each well containing 100 μl of culture medium with 10 ⁵ amastigotes from axecic culture with or without a serial drug dilution. The highest concentration for the test compounds is 90 μg/ml. Seven 3-fold dilutions are used covering a range from 30 μg/ml to 0.041 μg/ml. Each drug is tested in duplicate and each assay is repeated at least once. After 72hours of incubation the plates are inspected under an inverted microscope to assure growth of the controls and sterile conditions.

10µl of Alamar Blue (12.5 mg resazurin dissolved in 1L distilled water) are now added to each well and the plates incubated for another 2 hours. Then the plates are read with a Spectramax Gemini XS microplate fluorometer (Molecular Devices Cooperation, Sunnyvale, CA, USA) using an excitation wave length of 536 nm and an emission wave length of 588 nm.

Data are analysed using the microplate reader software Softmax Pro (Molecular Devices Cooperation, Sunnyvale, CA, USA).

Primary screen

The compounds are tested in duplicate at 7 concentrations. Miltefosine is used as the reference drug and shows an IC₅₀ value of 0.12 µg/ml.

If the IC $_{50}$ is >3 μ g/ml, the compound is classified as inactive

If the IC₅₀ is 0.1-3 μ g/ml, the compound is classified as moderately active lf the IC₅₀ is <0.1 μ g/ml, the compound is classified as active

Secondary screen

Active and moderately active compounds are tested in the macrophage assay with intracellular amastigotes in their host cells, murine macrophages.

Example B8

Biological Results

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Compounds of the invention, such as those in the examples above typically show activities in the low micromolar range for Plasmodium falciparum enzyme (K_i) and cell culture (ED_{50}), with selectivity (SI) over the human enzyme of at least 10-fold:

	R	Ki uM	SI	ED ₅₀ uM
O II	Ph ₃ CO	1.8	10	6
NH	TBDPSO	4.2	191	6.6
N O	TPSO	2.8	324	1.1
RO	Ph₃CNH	0.2	230	4.5
он				•
O	Ph ₃ CO	515	nd	1
ŅΗ	Ph ₃ NH	313	nd	1.8
- NOO				
ROJ				
ОТВОМЅ				

	Ki uM	SI	ED ₅₀ uM
TBDPSO	1.2	>833	3.0
TPSO	1.3	>769	1.0
TBDPSO	89	9	8.8
TPSO	975	nd	1.0
Ph ₃ CO	5	91	2.0
Ph₃NH	12	>83	5.3
	TBDPSO TPSO Ph ₃ CO	TPSO 1.3 TBDPSO 89 TPSO 975 Ph ₃ CO 5	TPSO 1.3 >769 TBDPSO 89 9 TPSO 975 nd Ph ₃ CO 5 91

Abbreviations

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TBDPSO	tert-butyldiphenylsilyloxy	DMF	dimethylformamide
TPSO	triphenylsilyloxy	DCM	dichloromethane
TBDMS	tert-butyldimethylsilyl	RT	room temperature
THF	tetrahydrofuran	Ac	acetyl
TEA	triethylamine	LAH	lithiumaluminiumhydride
TLC	thin layer chromatography	DMAP dimethylaminopyridine	

Throughout the specification and the claims which follow, unless the context requires otherwise, the word 'comprise', and variations such as 'comprises' and 'comprising', will be understood to imply the inclusion of a stated integer, step, group of integers or group of steps but not to the exclusion of any other integer, step, group of integers or group of steps.